

A new *Ypsilopus* (Orchidaceae, Angraecinae) from Zimbabwe and notes on the parallel evolution of extreme column exsertion in African angraecoids

João Farminhão^{1,2,3}, Phillip J. Cribb⁴

- 1 Jardim Botânico da Universidade de Coimbra, Coimbra, Portugal
- 2 Centre for Functional Ecology, Laboratório Associado TERRA, Departamento de Ciências da Vida, Universidade de Coimbra, Coimbra, Portugal
- 3 Herbarium et Bibliothèque de Botanique africaine, Université libre de Bruxelles, Brussels, Belgium
- 4 Herbarium, Library, Art & Archives, Royal Botanic Gardens, Kew, Richmond, UK

Corresponding author: João Farminhão (joao.farminhao@gmail.com)

Academic editor: Isabel Larridon + Received 31 May 2023 + Accepted 26 September 2023 + Published 19 October 2023

Abstract

Background and aims – A preliminary review of hawkmoth-pollinated angraecoids from Africa unveiled a remarkable case of parallel evolution of extreme column exsertion between the two species formerly classified in in the defunct genus *Barombia*. These belong to one clade of *Aerangis*, including *A. gracillima* and *A. stelligera*, and *Ypsilopus* sect. *Barombiella*, including *Y. amaniensis* and *Y. schliebenii*. The exploration of the geographical distribution of these two clades, followed by an examination of morphological variation within *Y. sect. Barombiella*, revealed that the disjunct population identified as *Y. amaniensis* from Zimbabwe represents an undescribed species.

Material and methods – Occurrence records of *Ypsilopus amaniensis*, *Y. schliebenii*, *Aerangis gracillima*, and *A. stelligera* were comprehensively mapped and distribution patterns were visually analysed. Pollination syndromes and pollinaria attachment sites were inferred based on a review of floral and hawkmoth morphology. Standard herbarium practices and mining of photographs of wild and cultivated plants in social media allowed the description of the novelty.

Key results – *Ypsilopus zimbabweensis* sp. nov. (*Y.* sect. *Barombiella*) is a narrow endemic of significant horticultural interest and it is preliminarily assessed as Endangered. The evolution of a *Barombia*-type column presents a parallel geographical pattern in the *Aerangis gracillima*-A. *stelligera* clade and *Ypsilopus* sect. *Barombiella* and probably induced a shift of pollen placement sites in these sphingophilous species.

Keywords

Great Zimbabwe National Monument, iNaturalist, lithophytic orchids, sphingophily, taxonomy, Tropical Africa

INTRODUCTION

The evolution of mechanical barriers leading to pollen placement shifts (i.e. changes of the position of pollen loads on pollinators) are exclusively reported in plants with high floral integration (i.e. coordinated covariance of floral traits) and accuracy (Armbruster and Muchhala 2009), notably in Orchidaceae (e.g. Dressler 1968; Nilsson et al. 1987). Column exsertion is one of the characters associated with the evolution of these shifts, as observed in Stylidiaceae (Armbruster et al. 1994) and Campanulaceae (Muchhala and Potts 2007; Muchhala 2008), but its importance in orchids remains undocumented.

A preliminary review of hawkmoth pollination in angraecoid orchids (Farminhão 2021), identified two clades of African species that possibly illustrate how column exsertion leads to shifts of pollen attachment sites in species sharing large hawkmoths as pollinators. In the Guineo-Congolian region, *Aerangis gracillima* (Kraenzl.) J.C.Arends & J.Stewart presents a column attaining more

Plant Ecology and Evolution is published by Meise Botanic Garden and Royal Botanical Society of Belgium.

Copyright João Farminhão, Phillip J. Cribb. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

than double the length of those of closely-allied *Aerangis stelligera* Summerh. and *A. bouarensis* Chiron. Similarly, in eastern and southeastern Africa, and with a more sizeable difference, *Ypsilopus schliebenii* (Mansf.) D'haijère & Stévart bears a column four to approximately five times longer than sister *Y. amaniensis* (Kraenzl.) D'haijère & Stévart. The striking resemblance in column elongation of *A. gracillima* and *Y. schliebenii* led taxonomists to class both species in the now defunct genus *Barombia* Schltr. (Schlechter 1914; Cribb 1980), which was revealed to be polyphyletic by molecular phylogenetics (Simo-Droissart et al. 2018; D'haijère et al. 2019; Farminhão et al. 2020).

To better understand this parallelism, we explored the geographical distribution of *A. stelligera*, *A. gracillima*, *Y. amaniensis*, and *Y. schliebenii* and examined in further detail morphological variation within these taxa, with a focus on the two species of *Ypsilopus* Summerh.

Ypsilopus is an angraecoid genus (from the cyrtridactyloid clade) confined to eastern and southern Africa, which encompasses 12 species arranged in two sections, comprising species formerly included in Tridactyle Schltr and Rangaeris (Schltr.) Summerh. (D'haijère et al. 2019, 2021; Farminhão et al. 2021). Ypsilopus schliebenii and Y. amaniensis are the only representatives of Ypsilopus sect. Barombiella (Szlach.) D'haijère & Stévart, which were formerly classified within polyphyletic Rangaeris (Farminhão et al. 2020). The wellknown Y. amaniensis is widespread in East Africa from Eritrea and Ethiopia south to Tanzania (Cribb 1989; Demissew et al. 2004). It is also reported from Zimbabwe, amongst others, by Ball (1978) and la Croix and Cribb (1998). However, the three collections (viz. Ball 1394, Jackson 56814, Mullin in GHS 25198) we have examined from there differ significantly from the East African ones, enough to consider them to belong to a distinct, albeit closely allied species which we describe here.

MATERIAL AND METHODS

We mapped all occurrences of *A. gracillima*, *A. stelligera*, *Y. amaniensis*, and *Y. schliebenii* vouchered by specimens deposited in BR, BRLU, P, K, and SRGH using QGIS v.3.4.15.

We compiled spur length and column measurements for the same angraecoid species based on a comprehensive literature survey (Cribb 1989; Geerinck 1992; la Croix and Cribb 1998; Szlachetko and Olsweski 2001). Body measurements of *Xanthopan morganii* (Walker, 1856), the largest hawkmoth from Tropical Africa, were estimated using ImageJ v.1.52d (Schneider et al. 2012) based on forewing length and iconography provided by Minet et al. (2021).

We applied standard herbarium practices to investigate the variability of plants identified as *Y. amaniensis* kept at BR, K, and SRGH, including reproductions of specimens kept at FI, and all type material (acronyms following Thiers 2023). Based on newly collected data, we update the key to the sections of *Ypsilopus*, with a focus on *Ypsilopus* sect *Barombiella*, presented by D'haijère et al. (2019, 2021). Distribution records and photographs of species in *Y*. sect. *Barombiella* were mined from social media, namely iNaturalist, Facebook, and Flickr, and used to complete the description, phenology, and range of the species.

IUCN Red List categories and criteria (IUCN 2022) were applied to evaluate the conservation status of the new species. The number of "locations" (sensu IUCN 2022) was calculated considering the type of threats, such that a single "location" may include more than one adjacent occurrence. The Extent of Occurrence (EOO) and Area of Occupancy (AOO) were calculated using GeoCAT (Bachman et al. 2011) on georeferenced specimen data. The AOO was calculated based on a 2×2 km grid cell size.

RESULTS

In *Aerangis* Rchb.f. and *Ypsilopus*, the *Barombia*-type column that has evolved in parallel occurs in the species with the narrowest distribution, which is marginal to the range of the most widespread taxa with shorter columns (Fig. 1). There is at least one contact zone between *A. gracillima* and *A. stelligera* in southern Cameroon, while species in *Y.* sect. *Barombiella* are separated by large geographical gaps: ca 200 km and 800 km separate *Y. schliebenii* from the closest populations of *Y. amaniensis* to the north and south of its range, respectively. The population of *Y. zimbabweensis*, newly described here, is separated by a gap of more than 1,200 km from the southernmost occurrence of *Y. amaniensis* in the Rubeho and Uluguru Mountains in Tanzania.

The range of spur and column length in the studied angraecoids is summarised in Table 1.

Ypsilopus zimbabweensis can be morphometrically separated from *Y. amaniensis* namely according to leaf length, flower number, and peduncle length. Differences are summarised in the taxonomic treatment.

DISCUSSION

Distribution of *A. gracillima*, *A. stelligera*, *Y. amaniensis*, and *Y. schliebenii* is consistent with different floristic bioregions of the Afrotropics (Droissart et al. 2018): in the Guineo-Congolian region, *A. gracillima* is confined to Lower Guinea, while sister *A. stelligera* is more widespread in Lower Guinea and Congolia; in Eastern Africa, the range of *Y. schliebenii* is centred in the Southern Rift montane region, whereas *Y. amaniensis* is more widespread in Central Tanzania and East African montane regions, with the isolated population of *Y. zimbabweensis* included in the South Zambesian. The geographical pattern of column length distribution suggests that column exsertion may have been reinforced after secondary contact in a scenario

	Spur length (cm)	Column length (cm)
Aerangis gracillima	18-25	3-4
Aerangis stelligera	14-25	1-1.5
Ypsilopus amaniensis	8-16	0.5
Ypsilopus schliebenii	16-17.5	2-2.7
Ypsilopus zimbabweensis	11-14	0.5

Table 1. Spur and column lengths in the Aerangis gracillima-A. stelligera clade and Ypsilopus sect. Barombiella.

of allopatric/peripatric speciation in both angraecoid clades (Farminhão 2021).

Considering spur length (see Johnson et al. 2017) and field observations (Martins and Johnson 2013; Balducci et al. 2019), it is likely that the *Aerangis gracillima–A. stelligera* clade and *Ypsilopus* sect. *Barombiella* share some large sphingid species as their exclusive pollinators, namely *Agrius convolvuli* (Linnaeus, 1758), *Coelonia fulvinotata* (Butler, 1875), and/or *Xanthopan morganii*. It is hypothesised that the pollinia attachment site shifted from the head to the dorsal region of the thorax (Fig. 1), based on the head-thorax length of 1.9 cm in *Xanthopan morganii*, mirroring the gap in column exsertion (see Table 1). The use of camera traps optimised for studying plant-insect interactions (e.g. Droissart et al. 2021) will be instrumental to test these hypotheses, notably near the contact zones between sister species. Populations previously identified as *Y. amaniensis* in Zimbabwe are here recognised as *Ypsilopus zimbabweensis* sp. nov. The novelty is apparently endemic to the Central Watershed biogeographical area (Mapaura 2002), and it is one of the five angiosperms restricted to the inselbergs of Zimbabwe (Seine et al. 1998; Mapaura 2002), the others being *Craterostigma syncerus* (Seine, Eb.Fisch. & Barthlott) Eb.Fisch., Schäferh. & Kai Müll., *Delosperma steytlerae* L.Bolus, *Kalanchoe wildii* Raym.-Hamet ex R.Fern., and *Portulaca rhodesiana* R.A.Dyer. The number of species in *Y.* sect. *Barombiella*, thus, rises to three, all presenting non-overlapping latitudinal distributions in Tropical Africa.

TAXONOMIC TREATMENT

Key to Ypsilopus

	sect. Ypsil	opus (see D'haijère et al. 2021)
2.	Column longer than 2 cm	Y. schliebenii
_	Column up to 2 cm long	
3.	Inflorescence 5–8-flowered; peduncle 1–1.5 cm	Y. amaniensis
_	Inflorescence 10–13-flowered; peduncle 4.5–7 cm	Y. zimbabweensis sp. nov.

Ypsilopus zimbabweensis Farminhão & P.J.Cribb, **sp. nov.**

urn:lsid:ipni.org:names:77328944-1 Figs 1-4

Type. ZIMBABWE • Masvingo [Victoria District], ± 3 km from Zimbabwe turn-off on Morgenster road; 12 Jan. 1976; *J.S. Ball 1394*; holotype: K; isotype: SRGH.

Diagnosis. Closely allied to *Ypsilopus amaniensis* (Kraenzl.) D'haijère & Stévart from eastern Africa but differs in having longer leaves (80–130 mm vs 35–115 mm in *Y. amaniensis*), inflorescences that greatly exceed the leaves, bearing 10–13 flowers (vs 5–8 in *Y. amaniensis*), and having a longer peduncle (45–70 mm vs 10–15 mm) and rachis (120–170 mm vs 50–80 mm).

Description. Robust, erect or rarely pendent, lithophytic or epiphytic <u>herb</u>, often forming clumps. <u>Roots</u> emerging through the leaf bases opposite the leaves, stout, 8–9 mm in diameter, branching distally, silvery grey. <u>Stems</u> 20–30

or more cm long, 7-9 mm in diameter, covered with sheathing leaf bases. Leaves rigidly coriaceous, 12-16, distichous, twisted just above the basal articulation to lie in one plane, linear-oblong, unequally roundly lobed at the apex, conduplicate at base just above the leaf sheath, $80-130 \times 12-19$ mm, deep olive-green, articulated to 10-17 mm long leaf sheath. Inflorescences longer than the leaves, arching to pendent, secund in two ranks, 1-several, from leaf sheaths 30–50 mm below the stem apex, 17–23 cm long, 10-13-flowered; peduncle cylindrical, 45-70 mm long, bearing 2-4 sheathing sterile bracts, 5-8 mm long; rachis slenderly cylindrical, slightly zigzag, 12-17 cm long; floral bracts cucullate, ovate, subacute, $6-8 \times$ 4–8 mm. Flowers 22×28 mm, showy, white with a bufftinged spur, the basal flower opening last, diurnally and nocturnally scented of vanilla; pedicel and ovary 22-25 mm long, the ovary scabrid. Sepals and petals reflexed at anthesis. Dorsal sepal linear-elliptic, acuminate, $15-20 \times$ 1.5-2 mm. Lateral sepals similar. Petals narrowly lineartapering, acuminate, $14-15 \times 1-1.5$ mm. Lip 3-lobed in the middle, $15-16 \times 5-6$ mm; side lobes obliquely oblong, truncate, $8-9 \times 2-3$ mm; midlobe linear-tapering, acuminate, 7-8 mm long; spur pendent, narrowly cylindrical from a narrow mouth, 110–140 mm long. <u>Column 5 mm long</u>, glandular; anther cap giving the tip of the column a hooked appearance; pollinia 2, stipes bifid with linear lobes; viscidium oblong. **Distribution.** Endemic to the Central Watershed of Zimbabwe, in the inselbergs of the southern middleveld margin of the Zimbabwe Craton, west of the Save River, in Masvingo Province (Fig. 1).

Habitat. Epiphyte or lithophyte on inselberg partly-shaded bare rock surfaces; 1000–1300 m.

Phenology. Flowers in the rainy season, from December to February.



Figure 1. Possible evidence for reinforcement in the geographical distribution and hypothetical pollinaria attachment sites (on a large sphingid hawkmoth) of two angraecoid clades with divergent column exsertion lengths in tropical Africa. *Aerangis gracillima* (yellow triangles) is closely allied to *A. stelligera* (blue triangles), while *Ypsilopus amaniensis* (blue diamonds) is closely related to *Y. schliebenii* (yellow diamonds). An extremely elongated *Barombia*-type column is present in *A. gracillima* and *Y. schliebenii*. The three isolated collections of *Y. amaniensis* in Zimbabwe are here assigned to *Ypsilopus zimbabweensis* (white diamonds). Photos by Murielle Simo-Droissart (*A. gracillima*), Bart Wursten (*A. stelligera*), Guido van Asten (*Y. amaniensis*), and Russell Hutton (*Y. schliebenii*).



Figure 2. *Ypsilopus zimbabweensis.* **A.** Habit. **B.** Flower, side view. **C, D.** Flower, front view. **E.** Lip margin variability. **F.** Column, ventral view, with glandular trichomes visible. **G.** Anther cap, side, dorsal, and ventral views. **H.** Viscidium and stipes. **I.** Pollinium (one of two). A (in part), E (in part), F–I drawn from the type collection; A, E (both in part) and I from *Jackson 56814*; B, C after watercolour by Patricia van de Ruit. All drawn by Andrew Brown.



Figure 3. Watercolour of *Ypsilopus zimbabweensis*, originally identified as *Rangaeris amaniensis*, by Patricia van de Ruit, published in Ball (1978: 1394). Reproduced with permission.

Etymology. The species is only recorded from Zimbabwe, namely from the area around the Great Zimbabwe National Monument, which gives the country its name.

Additional material (paratypes). ZIMBABWE . Masvingo [Victoria District], Mt Morgenster; 1000 m (3500 ft); fl. in cult. Harare [Salisbury]; 24 Jan. 1956; R.W. Jackson 56814; K!, SRGH • Masvingo, 16 km NW of Ndanga; 29 Dec. 1976; L.J. Mullin in GHS 25198; SRGH!. Preliminary IUCN conservation assessment. The species is given a Red List status of Endangered: EN B1ab(v)+B2ab(v). Ypsilopus zimbabweensis is known from three collections and one observation (https://www. inaturalist.org/observations/143791156) made between 1956 and 2012, representing four occurrences and three locations, including one within the Great Zimbabwe National Monument, a Cultural World Heritage Site. The extent of occurrence (EOO) is 132.1 km² and the area of occupancy (AOO) is 16 km². The EOO and AOO fall within the limits of the Endangered (CR) category under subcriteria B1 and B2. Since this species occurs only in three locations and a decline of mature individuals is projected because of illegal collection for the orchid trade, it meets condition b(v) for the EN category.

Notes. *Ypsilopus amaniensis* is to be excluded from Flora Zambesiaca, since all regional occurrences correspond to *Y. zimbabweensis*, namely the recent records illustrated on the Flora of Zimbabwe website (Hyde et al. 2023). Specimens identified as *Angraecum* sp. in the inselbergs of Zimbabwe (Seine et al. 1998) are also possibly ascribable to *Y. zimbabweensis*. Patricia van de Ruit's fine watercolour

illustration of this species (as *Rangaeris amaniensis*) is reproduced here (Fig. 3). Iconography produced by the same artist, for the same book, was instrumental to the description of another new orchid from Zimbabwe (Farminhão and Cribb 2020). The new species is also illustrated here with a line drawing by Andrew Brown (Fig. 2) and a photograph in la Croix and la Croix (1997), reproduced here (Fig. 3). The novelty has been widely cultivated and misidentified as *Rangaeris amaniensis* by hobbyists in Zimbabwe, South Africa, Australia, and Europe.

ACKNOWLEDGEMENTS

We thank the curators and staff of BR, BRLU, K, P, and SRGH for making their collections available and for kindly allowing the authors to use the facilities of their institutions. We thank Andrew Brown for the excellent line drawing of the new species and Patricia van de Ruit for her watercolour, the latter reproduced with permission of Jane Browning, the late John Ball's sister. We thank Bart Wursten, Guido van Asten, Isobyl la Croix, and Russell Hutton for allowing us to use their photographs of *Ypsilopus* sect. *Barombiella*. We are grateful to Nicolas Texier for assisting us with the production of the distribution map. Herbarium visits of the first author were supported by the Belgian Fund for Scientific Research (F.R.S-FNRS). The first author's PhD research was funded by the Belgian Fund for Research Training in Industry



Figure 4. *Ypsilopus zimbabweensis.* **A.** Plants growing as lithophytes in situ. **B.** Inflorescence, side view, of plant cultivated in Harare. Photos by Bart Wursten (A) and Isobyl la Croix (B).

and Agriculture (FRIA) of the F.R.S-FNRS (scholarships F 3/5/5 – FRIA/FC 33848881) and by the Van Buuren-Jaumotte-Demoulin Prize, awarded by the David and Alice Van Buuren Fund. An earlier version of this article is part of J.F.'s PhD thesis entitled 'Advances in angraecoid orchid systematics in Tropical Africa and Madagascar: new taxa and hypotheses for their diversification' defended at the Université libre de Bruxelles in 2021. The first author thanks Tariq Stévart and Pierre Meerts for supervising his PhD work. Finally, we are grateful to Benny Bytebier, an anonymous reviewer, and Isabel Larridon for their corrections on an earlier version of this manuscript.

REFERENCES

- Armbruster WS, Edwards ME, Debevec EM (1994) Floral character displacement generates assemblage structure of Western Australian triggerplants (*Stylidium*). Ecology 75: 315–329. https://doi.org/10.2307/1939537
- Armbruster WS, Muchhala N (2009) Associations between floral specialization and species diversity: cause, effect, or correlation? Evolutionary Ecology 23: 159–179. https://doi. org/10.1007/s10682-008-9259-z
- Bachman S, Moat J, Hill AW, De La Torre, J, Scott B (2011) Supporting Red List threat assessments with GeoCAT: geospatial conservation assessment tool. ZooKeys 150: 117– 126. https://doi.org/10.3897/zookeys.150.2109
- Balducci MG, Martins DJ, Johnson SD (2019) Pollination of the long-spurred African terrestrial orchid *Bonatea steudneri* by long-tongued hawkmoths, notably *Xanthopan morganii*. Plant Systematics and Evolution 305: 765–775. https://doi. org/10.1007/s00606-019-01605-2
- Ball JS (1978) Southern African Epiphytic Orchids. Conservation Press (Pty) Ltd, Johannesburg & London, 1–247.
- Cribb PJ (1980) *Barombia schliebenii* an extraordinary epiphytic orchid from central Tanzania. South African Orchid Journal 11: 110.
- Cribb PJ (1989) *Rangaeris*. In: Polhill R (Ed.) Flora of Tropical East Africa. Orchidaceae (Part 3). A.A. Balkema, Rotterdam, 570–573.
- Demissew S, Cribb P, Rasmussen FN (2004) Field Guide to Ethiopian Orchids. Royal Botanic Gardens, Kew, Richmond, 1–304.
- D'haijère T, Mardulyn P, Dong L, Plunkett GM, Simo-Droissart M, Droissart V, Stévart T (2019) Molecular phylogeny and taxonomic synopsis of the angraecoid genus *Ypsilopus* (Orchidaceae, Vandeae). Taxon 68: 455–470. https://doi.org/10.1002/tax.12072
- D'haijère T, Farminhão JNM, Stévart T, Fischer E (2021) Novelties among East African angraecoids (Orchidaceae, Angraecinae). Nordic Journal of Botany 39: e03184. https:// doi.org/10.1111/njb.03184
- Dressler RL (1968) Pollination by euglossine bees. Evolution 22: 202–210. https://doi.org/10.2307/2406664
- Droissart V, Dauby G, Hardy OJ, Deblauwe V, Harris DJ, Janssens S, Mackinder BA, Blach-Overgaard A, Sonké B, Sosef MM, Stévart T, Svenning J-C, Wieringa JJ, Couvreur

- Droissart V, Azandi L, Onguene ER, Savignac M, Smith TB, Deblauwe V (2021) PICT: a low-cost, modular, open-source camera trap system to study plant-insect interactions. Methods in Ecology and Evolution 12: 1389–1396. https:// doi.org/10.1111/2041-210X.13618
- Farminhão J (2021) Advances in angraecoid orchid systematics in Tropical Africa and Madagascar: new taxa and hypotheses for their diversification. PhD Thesis, Université libre de Bruxelles, Belgium.
- Farminhão J, Cribb PJ (2020) Two new species of *Rhipidoglossum* (Orchidaceae: Angraecinae) from Tanzania and Zimbabwe. Kew Bulletin 75: 30. https://doi.org/10.1007/ s12225-020-09888-2
- Farminhão JNM, D'haijère T, Droissart V, Dumbo Isonga L, Dong L, Verlynde S, Plunkett GM, Simo-Droissart, M, Stévart T (2020) An elegy to *Rangaeris*, including a description of two new genera in the *Cyrtorchis-Tridactyle* clade (Orchidaceae, Angraecinae). Annals of the Missouri Botanical Garden 105: 300–322. https://doi. org/10.3417/2020472
- Farminhão JNM, Verlynde S, Kaymak E, Droissart V, Simo-Droissart M, Collobert G, Martos F, Stévart T (2021) Rapid radiation of angraecoids (Orchidaceae, Angraecinae) in Tropical Africa characterised by multiple karyotypic shifts under major environmental instability. Molecular Phylogenetics and Evolution: 107105. https://doi. org/10.1016/j.ympev.2021.107105
- Geerinck D (1992) Flore d'Afrique Centrale (Zaïre-Rwanda-Burundi): Spermatophytes; Orchidaceae (seconde partie). Jardin Botanique National de Belgique, Meise, 297–780. https://doi.org/10.5281/zenodo.4660246
- Hyde MA, Wursten BT, Ballings P, Coates Palgrave M (2023) Flora of Zimbabwe. Species information: *Rangaeris amaniensis*. https://www.zimbabweflora.co.zw/speciesdata/ species.php?species_id=118930 [accessed 15.09.2023]
- IUCN (2022) Guidelines for Using the IUCN Red List Categories and Criteria. Version 15. https://www.iucnredlist. org/resources/redlistguidelines [accessed 03.04.2023]
- Johnson SD, Moré M, Amorim FW, Haber WA, Frankie GW, Stanley DA, Cocucci AA, Raguso RA (2017) The long and the short of it: a global analysis of hawkmoth pollination niches and interaction networks. Functional Ecology 31: 101–115. https://doi.org/10.1111/1365-2435.12753
- la Croix I, Cribb PJ (1998) *Rangaeris*. In: Pope G (Ed.) Flora Zambesiaca 11(2). Orchidaceae. Royal Botanic Gardens, Kew & Flora Zambesiaca Managing Committee, London, 438–442.
- la Croix I, la Croix E (1997) African Orchids in the Wild and in Cultivation. Timber Press, Portland, Oregon, 1–379.
- Mapaura A (2002) Endemic plant species of Zimbabwe. Kirkia 18: 117–149. https://www.jstor.org/stable/23502383
- Martins DJ, Johnson SD (2013) Interactions between hawkmoths and flowering plants in East Africa: polyphagy and evolutionary specialization in an ecological context.

Biological Journal of the Linnean Society 110: 199-213. https://doi.org/10.1111/bij.12107

- Minet J, Basquin P, Haxaire J, Lees DC, Rougerie R (2021) A new taxonomic status for Darwin's "predicted" pollinator: *Xanthopan praedicta* stat. nov. Antenor 8: 69–86. http://doi. org/10.5281/zenodo.5856141
- Muchhala N (2008) Functional significance of interspecific variation in *Burmeistera* flower morphology: evidence from nectar bat captures in Ecuador. Biotropica 40: 332–337. https://doi.org/10.1111/j.1744-7429.2007.00381.x
- Muchhala N, Potts MD (2007) Character displacement among bat-pollinated flowers of the genus *Burmeistera*: analysis of mechanism, process and pattern. Proceedings of the Royal Society B 274: 2731–2737. https://doi.org/10.1098/ rspb.2007.0670
- Nilsson LA, Johnsson L, Ralison L, Randrianjohany E (1987) Angraecoid orchids and hawkmoths in central Madagascar: specialized pollination systems and generalist foragers. Biotropica 19: 310–318. https://doi.org/10.2307/2388628
- Schlechter R (1914) Die Orchideen: ihre Beschreibung, Kultur und Züchtung. P. Parey, Berlin, 1–837. https://doi. org/10.5962/bhl.title.46623

- Schneider CA, Rasband WS, Eliceiri KW (2012) NIH Image to ImageJ: 25 years of image analysis. Nature Methods 9: 671– 675. https://doi.org/10.1038/nmeth.2089
- Seine R, Becker U, Porembski S, Follmann G, Barthlott W (1998) Vegetation of inselbergs in Zimbabwe. Edinburgh Journal of Botany 55(2): 267–293. https://doi.org/10.1017/ S0960428600002195
- Simo-Droissart M, Plunkett GM, Droissart V, Edwards MB, Farminhão JNM, Ječmenica V, D'haijère T, Lowry II PP, Sonké B, Micheneau C, Carlsward BS, Azandi L, Verlynde S, Hardy OJ, Martos F, Bytebier B, Fischer E, Stévart T (2018) New phylogenetic insights toward developing a natural generic classification of African angraecoid orchids (Vandeae, Orchidaceae). Molecular Phylogenetics and Evolution 126: 241–249. https://doi.org/10.1016/j. ympev.2018.04.021
- Szlachetko DL, Olszewski TS (2001) Orchidacées 3. In: Achoundong G, Morat P (Eds) Flore du Cameroun 36. Ministère de la Recherche Scientifique et Technique, Yaoundé, 666–948.
- Thiers B (2023) Index Herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. https://sweetgum.nybg.org/science/ih/ [accessed 27.09.2023]