

Keetia nodulosa sp. nov. (Rubiaceae - Vanguerieae) of West - Central Africa: bacterial leaf nodulation discovered in a fourth genus and tribe of Rubiaceae.

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Summary. *Keetia nodulosa* Cheek, a cloud forest climber nearly endemic to Cameroon, with a single record from Nigeria, is described and illustrated. It is remarkable as the first known species to be recorded with bacterial leaf nodules (BLN) in the genus *Keetia*, and also, in the tribe Vanguerieae. Other genera in Rubiaceae with BLN are *Psychotria* (Psychotrieae), *Sericanthe* (Coffeaeae) and *Pavetta* (Pavetteae). The BLN in *Keetia* (Vanguerieae) are illustrated for the first time here.

The characteristics and significance of bacterial leaf nodulation in *Keetia nodulosa* are discussed in the context of rapidly growing knowledge on the subject in flowering plants. *Keetia nodulosa* is provisionally assessed using the 2012 IUCN standard as Endangered (EN B2ab(iii)). The importance of its conservation, and options for achieving this are discussed in the context of recent extinctions of other plant species in Cameroon.

This discovery of a new cloud forest species is discussed in relation to other cloud forest plant species described in the last twenty years which are also distributed over the highlands of the western half of Cameroon.

Key words. Bacterial leaf nodules; *Burkholderia*; cloud forest; conservation; horizontal transfer; *Paraburkholderia*

Introduction

Keetia S.M. Phillips was segregated from *Canthium* Lam. by Bridson (1985, 1986). Restricted to sub-Saharan Africa, and extending from Guinea in West Africa (Gosline *et al.* 2023a; 2023b) to Sudan in the North and East (Darbyshire *et al.* 2015) and S. Africa in the South (Bridson 1986), this genus of about 40 accepted species (POWO, continuously updated) are mainly forest climbers, distinguished from similar Canthioid genera in west Africa by their pyrenes with a fully or partly-defined lid-like area around a central crest (Bridson 1986). In a phylogenetic analysis of the tribe based on morphology, nuclear ribosomal ITS and chloroplast *trnT-F* sequences, Lantz & Bremer (2004), found that based on a sample of four species, *Keetia* was monophyletic and sister to *Afrocanthium* (Bridson) Lantz & B. Bremer with strong support. Highest species diversity of *Keetia* is found in

Cameroon and Tanzania, both of which have about 15 taxa (Onana 2011; POWO, continuously updated). In contrast, neighbouring Gabon has only 10 species, although most specimens recorded remain unidentified to species, Sosef *et al.* 2006). Several *Keetia* species are point endemics, or rare national endemics, and have been prioritized for conservation (e.g. Onana & Cheek 2011; Couch *et al.* 2019; Murphy *et al.* 2023; Darbyshire *et al.* 2023) and one threatened species, *Keetia susu* Cheek has a dedicated conservation action plan (Couch *et al.* 2022)

Bridson's (1986) account of *Keetia* was preparatory to treatments of the Vanguerieae for the Flora of Tropical East Africa (Bridson & Verdcourt 1991) and Flora Zambesiaca (Bridson 1998). Pressed to deliver these, she stated that she could not dedicate sufficient time to a comprehensive revision of the species of *Keetia* outside these areas: "full revision of *Keetia* for the whole of Africa was not possible because the large number of taxa involved in West Africa, the Congo basin and Angola and the complex nature of some species would have caused an unacceptable delay in completion of some of the above Floras" (Bridson 1986). Further "A large number of new species remain to be described." Several of these new species were indicated by Bridson (1986), and other new species by her arrangement of specimens in folders that she annotated in the Kew Herbarium. One of these species was later taken up and published by Jongkind (2002) as *Keetia bridsoniae* Jongkind. In the same paper, Jongkind discovered and published *Keetia obovata* Jongkind based on material not seen by Bridson. Based mainly on new material, additional new species of *Keetia* have been published by Bridson & Robbrecht (1993), Bridson (1994), Cheek (2006), Lachenaud *et al.* (2017), Cheek *et al.* (2018a) and Cheek & Bridson (2019).

In the course of formally publishing new species to science from Cameroon so that they could be Red Listed and considered for inclusion in the Cameroon Important Plant Areas programme (e.g. Murphy *et al.* 2023), numerous new species to science have been published (see below), mainly based on species informally identified as new in the course of a series of surveys for improved conservation management of plant species and habitats conducted mainly in western Cameroon in the 1990s (Cheek *et al.* 2006). This paper continues the endeavor.

In this paper, a remarkable new species of *Keetia*, *K. nodulosa* Cheek is described. *Keetia nodulosa* is unique in its genus and tribe for having conspicuous bacterial nodules on its abaxial leaf blade surfaces, resembling those seen in species of the genus *Pavetta* L., which also have conspicuous black nodules at nerve junctions. The presence of bacterial nodules was first reported in the conservation checklist "The Plants of Mount Kupe, Muanenguba and the Bakossi Mts" (Cheek *et al.* 2004: 375). The specimens Etuge 2798 and Etuge 2829 (both Mt Kupe) were matched with specimens from Cameroon, that had been included in the protologue of *Keetia purseglovei* Bridson (Bridson 1986), Zenker 2986 (Bipinde) and Zenker & Staudt 415 (Yaoundé). However, the two Etuge specimens concerned had been annotated as "vel sp. aff.", indicating that they might represent another but related species. Further research showed that all the Ugandan material of *Keetia purseglovei*, including the type, lacked bacterial nodules, and while and filed similar to the Cameroonian material, differed in several morphological characters (see Table 1 below). In searching all other material of *Keetia* at K, and other herbaria, for bacterial nodules, an additional specimen, *Emwiogbon* FHI 65823 from Nigeria, close to the Cameroon border, was found. This matched the Cameroonian material of *K. nodulosa*. It had been identified as a second specimen of *Keetia inaequilatera* (Hutch. & Dalz.) Bridson. While similar to the type and only other known specimen of that species, characters were found that separated this

specimen from the type of that species (see Table 1 and diagnosis below) including the presence (vs absence) of bacterial nodules. Finally, just before the paper was completed, a further specimen, with flower buds, *Gereau et al.* 5639 from the Rumpi Hills, that had been identified as *K. cf. hispida*, was encountered and also placed in *K. nodulosa* in view of having bacterial nodules and other concordant characteristics.

Further searches on gbif.org revealed that additional specimens had been identified as *Keetia purseglovei*, mainly from Gabon, Central African Republic, R.D. Congo and Congo-Brazzaville. However, these differed from *K. nodulosa*, and only one of these, *Texier* 2164, possessed visible bacterial nodules (see notes below) so were discounted.

In this paper it is shown that two specimens from Cameroon previously ascribed to *Keetia purseglovei* in Bridson (1986) together with additional specimens, are specifically distinct from the Ugandan material of that species, including the type. The Cameroonian taxon, which extends to Nigeria, is formally characterized and named as *Keetia nodulosa* sp. nov.

Within Africa, Cameroon remains a major source of discovery for new species to science of vascular plants, with more species new to science published per annum than any other country in tropical Africa (Cheek *et al.* 2020a). Recent novelties range from forest trees (Quintanar *et al.* 2023; Cheek *et al.* 2022a; 2023a), shrubs and small trees (Couvreux *et al.* 2022; Gosline *et al.* 2022; Stone *et al.* 2023; Cheek *et al.* 2023b), lianas (Jongkind & Lachenaud 2022), rheophytes (Cheek *et al.* 2022b), terrestrial herbs (Cheek *et al.* 2021), to epilithic herbs (Janssens *et al.* 2022; Cheek *et al.* 2023c) and ferns (Shang & Zhang 2023; Dubuisson *et al.* 2022).

Materials and Methods

Names of species and authors follow IPNI (continuously updated). Herbarium material was collected using the patrol method e.g. Cheek & Cable (1997). Identification and naming follows Cheek in Davies *et al.* (2023). Herbarium specimens were examined with a Leica Wild M8 dissecting binocular microscope fitted with an eyepiece graticule measuring in units of 0.025 mm at maximum magnification. The drawing was made with the same equipment with a Leica 308700 camera lucida attachment. Pyrenes were characterized by boiling selected ripe fruits for several minutes in water until the flesh softened and could be removed. Finally, a toothbrush was used to clean the exposed pyrene surface to expose the surface sculpture. Specimens were inspected from the following herbaria: BM, BR, K, P, WAG, YA. It was not possible to view the duplicates of *Keetia nodulosa* deposited at YA because they are thought to be in the mounting backlog (Onana pers. obs. Feb. 2024). The format of the description follows those in other papers describing new species of *Keetia*, e.g. Cheek & Bridson (2019). Terminology follows Beentje & Cheek (2013). Herbarium codes follow Index Herbariorum (Thiers, continuously updated). Nomenclature follows Turland *et al.* (2018). All specimens seen are indicated “!” The conservation assessment follows the IUCN (2012) standard.

Results

Table 1. Characters distinguishing *Keetia inaequilatera*, *K. nodulosa* sp.nov. and *Keetia purseglovei*. Data for the first and third species from Bridson (1986) and specimens at K.

	<i>Keetia inaequilatera</i>	<i>Keetia nodulosa</i> sp. nov.	<i>Keetia purseglovei</i>
Distribution	S.E. Nigeria	S.E. Nigeria & Cameroon	Uganda (and probably eastern DR Congo)
Habitat	Lowland forest <800 m alt.	Cloud forest 800 – 940 m alt.	Submontane forest 1200 – 1265 m alt.
Secondary (spur) shoots, number of nodes	(2 –)3 – 4(– 5)	7 – 10(– 13)	3 – 5
Bacterial nodules on abaxial surface of leaf blades	Absent	Conspicuous along tertiary nerves	Absent
Domatia, position	On the secondary nerve bases	In the axils of the secondary nerves	In the axils of the secondary nerves
Domatia, number of hairs	10 – 30	10 – 30	0 (–5)
Stipule persistence (fruiting stage)	Unknown	Highly caducous, present usually only at the distalmost node	Persistent for 3 – 4 nodes from the apex
Stipule blade shape at maturity	Triangular	Subquadrate	Transversely elliptic
Pedicel length (mm)	2 – 3	(1.8 –)2.5 – 3(–4)	5 – 7
Flower bud shape and surface	Capitate (apex ovoid, base narrow cylindrical); papillate	Clavate (obovoid) to capitate; smooth	Constricted (waisted) in middle to capitate; smooth
Calyx indumentum	Glabrous	Apex of teeth densely long hairy (rarely glabrous)	Glabrous, or teeth with 2–3 hairs at apex
Leaf-blade shape (proximal leaves of spur shoots) and length: breadth ratio	Broadly ovate to suborbicular 1.2 – 1.5:1	Narrow elliptic or obovate-elliptic (2 –)3: 1	Elliptic (rarely oblanceolate-elliptic) 2 – 2.5:1
Leaf-blade colour, abaxial surface	Mid to dark brown	Grey-black (rarely green)	Pale orange or pale green
Number of secondary nerves on each side of the midrib	(3 –)4(– 5)	5 – 7	(4 –)5 – 6

Taxonomic treatment

***Keetia nodulosa* Cheek sp. nov.** Type: Cameroon, S.W. Province [now Region], Kupe-Muanenguba Division, alt. 850 m, Kupe Village, main trail towards Mount Kupe, forest near

a valley, fr.16 July 1996, *Etuge* 2798 with Felix, Ewang, Bishop, P., Temple, R. (holotype K000109898!; isotypes BR0000025613452V!, MO, P, WAG1966136!, YA).

LSID: urn:lsid:ipni.org:names:77336635-1

Keetia purseglovei Bridson (1986: 972) *quoad* Zenker 2986 (BM!, BR!, K!, P!) and Zenker & Staudt 415 K!); Cheek *et al.* (2004: 375).

Evergreen climber, climbing with clasping fruiting peduncles, 5 – 10 m tall. Primary stems with distal internodes glabrous, drying purple at first, subglossy, longitudinally finely ridged (microscope needed), distal internode flattened, other internodes subterete with a small central hollow, 5.3 – 6.5 x 0.35 – 0.4 cm, (distal, fertile internodes) at length with epidermis becoming longitudinally streaked with white. Secondary shoots (brachyblasts, plagiotropic or spur shoots) leafy, opposite, subequal in pairs, each 12– 37 cm long, with 4 – 9 internodes, internodes 2.5 – 6.1 x 0.12 – 0.2.5(– 0.3) cm, otherwise as the primary stems (Fig. 1 A), glabrous at fruiting stage, at flowering stage with sparse, patent, bristle hairs as the leaves. Leaves of primary axis not seen; those of secondary shoots distichous, not dimorphic, opposite and equal at each node, thinly leathery to thickly papery, blades drying black on upper surface, grey-black, rarely grey-green, on lower surface, elliptic, narrowly elliptic, or obovate elliptic, (4.9 –)5.3– 8.5(– 10.8) x 2.3 – 3.9(– 4.7) cm, acumen triangular 0.4 – 1.1(– 1.5) x 0.25 – 0.5 cm long, apex rounded; base obtuse to broadly acute, or rounded, rarely subcordate, usually asymmetric and decurrent on petiole; midrib and secondary nerves dull white to pale yellow, raised on both surfaces; domatia pit-like, longitudinally elliptic-oblong, 0.4 – 0.55 x 0.25 mm, inserted in the axil of midrib and the subtending secondary nerve, with 14 – 25 copper-coloured bristle hairs c. 0.1 mm long inserted around the rim, directed randomly: inconspicuous on upper surface; margin slightly thickened, revolute; secondary nerves 5 – 6(– 7) on each side of the midrib, arising at 50– 60° from the midrib, curving gradually upwards, the apex terminating parallel to and 3 – 4 mm from the blade margin, sometimes uniting with the nerve above. Tertiary nerves faintly visible, quaternary nerves inconspicuous. Bacterial nodules conspicuous on the abaxial surface, jet black, mainly at the junctions or along the lengths of tertiary nerves, about (1 –)2 –4(–5) mm apart, each 0.75 –2.5 mm long, usually with 3 –7 short lateral lobes along their length, 0.3 mm (unlobed) or 0.5 –0.75 mm wide, with a few smaller, unlobed, T-shaped or comma shaped nodules interspersed (Fig. 1C); hairs sparse, 3 – 20 % cover, along the midrib, secondary nerves (abaxial surface), and margins, simple, pale bronze-coloured, 0.25 – 0.5 mm long, strigose, slightly curved from base to apex, distal part gradually tapering to an acute apex, leaf otherwise glabrous. Petioles plano-convex in transverse section, the adaxial surface extended as narrow wings, (0.4 –) 0.5 – 0.8(– 0.9) x 0.1 cm, indumentum as midrib of blade. Stipules free, caducous (usually persisting at terminal node only at fruiting stage), glabrous apart from colleters, at apical bud narrowly triangular, c. 8 x 2 mm, the blade not distinct from the awn; at older nodes (flowering stage only, Fig. 1D & E) the blade distinct, subquadrate, widest at base, 4.5 –5 x 4.5 –5 mm, midrib not conspicuous; awn excurrent from the outer surface of the bade, arising below the apex, c. 6 x 0.8 – 1 mm, terete (or longitudinally 5 ridged on both

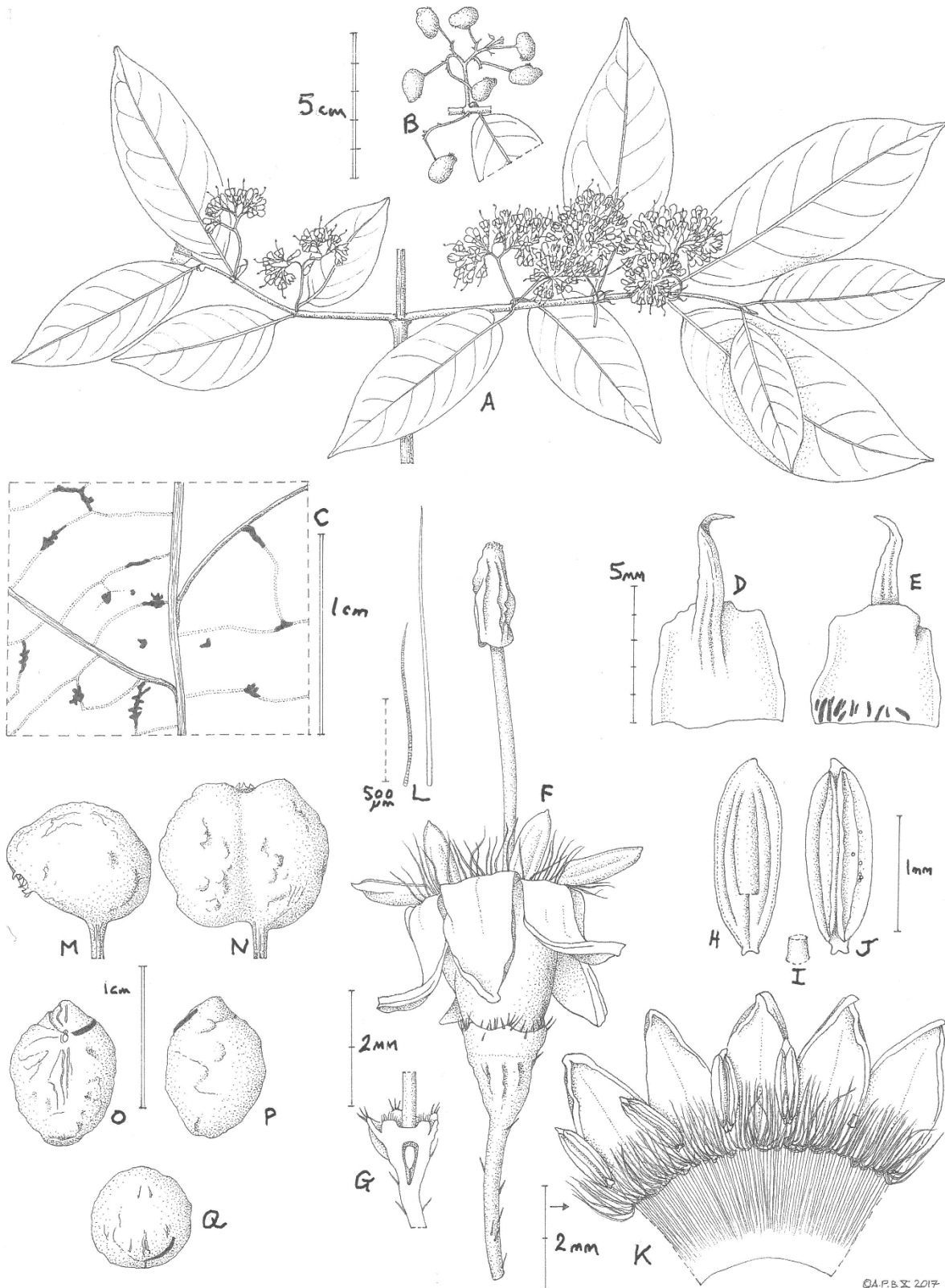


Fig. 1. *Keetia nodulosa*. **A.** habit, flowering secondary (short plagiotropic or spur) shoots; **B.** infructescence; **C.** leaf-blade, abaxial surface showing bacterial nodules; **D.** stipule abaxial (outer) surface; **E.** stipule adaxial (inner) surface showing colleters; **F.** flower; **G.** near longitudinal section of flower base, showing disc; **H.** anther, outer surface; **I.** filament; **J.** anther, inner face; **K.** corolla, opened (one stamen removed); **L.** moniliform (L) and bristle (R) hairs from inner surface of corolla tube (see K); **M.** single seeded fruit; **N.** double seeded fruit;

O pyrene, frontal view; **P** pyrene, side view; **Q** pyrene, plan view. **A, C-L** from *Zenker* 415; **B, M-Q** from *Etuge* 2798. Drawn by ANDREW BROWN.

surfaces), apex acute; colleters inserted on the adaxial surface near the base, botuliform, 0.4 – 0.8 x 0.2 mm, apex rounded. *Inflorescences* (Fig. 1A), axillary on spur (plagiotropic) branches, held above the stem, in 2 – 4 successive nodes beginning 1– (– 2) nodes below stem apex; anthesis ± simultaneously at all nodes, each inflorescence (11–) 40 – 60-flowered, forming heads 2.8 – 4.8 x 1.2 – 1.5 cm. Peduncles 15 – 22 x 0.75 mm, with two pairs of bracts 4 – 5 mm below the apex, bracts triangular c. 0.75 x 0.4 mm, membranous, sparsely and inconspicuously simple hairy; branches two, equal, 4 – 7 mm long, with each branch further forked, or terminating in a fascicle of 5 – 12 flowers. *Pedicels* 2.5 – 3(–4) x 0.75 mm long, with several scattered, slightly spreading, straight, acute hairs 0.25 mm long. Calyx-hypanthium obconical 1 x 1.25 mm, with c. 5 shallow longitudinal grooves, calyx tube shortly cylindrical, 0.3 – 0.4 mm long; teeth 5, very shortly and broadly triangular, 0.1– 0.2 x 0.5 mm, the margins of the teeth apices with dense erect, simple hairs 0.1 – 0.15 mm long as the pedicel, rarely absent, or, with a few on the abaxial surface (Fig. 1F&G). Corolla in bud clavate or narrowly obovoid, unconstricted 4.2 – 4.5(– 5) x 2 – 2.5 mm, apex rounded; at anthesis white, tube 3 x 2 mm, lobes 5, valvate, reflexed, oblong triangular, 2 x 1 – 1.5 mm, mouth with exserted, moniliform white hairs 0.7 – 1.5 mm long, from a ring inserted 0.3 – 0.4 mm below the mouth and 2 mm above the base of the corolla tube (Fig. 1F&K); inner surface glabrous from base to a ring of translucent deflexed bristle hairs c. 1.5 mm long adjacent to the ring of exserted hairs, inserted c. 2 mm above base (Fig. 1K). Stamens 5, inserted just below the corolla tube mouth, erect, filaments flat, 0.2 – 0.3 x 0.2 mm (Fig. 1I); anthers exserted, introrse, narrowly ellipsoid, 1.5 x 0.5 mm, apical connective appendage conical, c. 0.1 – 0.1 mm (Fig. 1H&J), sub-basifixed, base minutely hastate, the two bases conical, splayed, c. 0.1 mm long, acute. Disc annular, truncate, c. 0.2 x 0.8 mm, puberulent, hairs c. 0.1 mm long (Fig. 1G). Style c. 9 mm long, 0.2 mm wide, terete, the apex with a narrowly cylindrical, 10-fluted head or *receptaculum pollinis*, c. 1.75 x 0.75 mm, stigmatic apex papillate. *Infructescences* 3 – 7(– 9)-fruited, peduncles, clasping, reflexing, axes glabrescent, with a few thinly scattered simple hairs 1 mm long (Fig. 1F.). *Fruit* green (mature fruits), fleshy, didymous, in side view suborbicular, 10 (– 11) x 12 (– 13) x 11 – 13 x 8 mm, the two carpels united along their length but divided by a shallow longitudinal groove on each side (Fig. 1N), apex shallowly retuse, apical sinus c. 1 x 5 mm, including calyx 2 mm diam., teeth persistent, disc inconspicuous (Fig. 1G.); base slightly cordate or rounded, surface with 2– 8 raised verrucae mainly on each side of each carpel, verrucae c. 1 x 1 mm; 1-seeded fruits (by abortion, the majority, 7/8 of all ripe fruit), ovoid-elliptic, asymmetric, (8 –)10– 11 x (7–)7.5– 8 mm. Pyrene pale brown, woody, subellipsoid, 0.9 – 1 x 0.7– 0.75 x 0.5 – 0.7 cm, the surface with low, irregular, orbicular raised areas c. 1 mm diam., interlaced with white fibres. Lid apical, cap-like, c. 2–3 x 6 x 6 mm, angled c. 20 degrees towards the ventral face, crest (keel) distinct, broad; ventral face sometimes with a transverse slit opening 2(–3) mm long, at junction with main body of pyrene. Fig. 1A-Q.

RECOGNITION. *Keetia nodulosa* differs from all known species of the genus in having bacterial nodules on the abaxial leaf blade surfaces (vs absent), further differing also from the similar *Keetia purseglovei* Bridson in the primary axis subterete (vs 4-fluted); stipules caducous at fruiting stage, persisting usually only at stem apex (vs persisting for 3 to 4 nodes from apex); stipule blades subquadrate (vs transversely elliptic); pedicels 2.5 – 3(– 4) mm long (vs 5 – 7 mm) From *K. inaequilatera* (Hutch.) Bridson differing in the narrow elliptic or obovate-elliptic leaf blades with length: breadth ratio (2 –)3: 1 (vs broadly ovate to suborbicular, 1.2 – 1.5:1), the domatia situated in the axils of the secondary nerves (vs on the

secondary nerve bases) and the flower bud smooth, (not with the corolla bud head minutely papillate). See Table 1 above for additional diagnostic characters.

DISTRIBUTION. S.E. Nigeria and Cameroon

SPECIMENS EXAMINED. NIGERIA. South-Eastern State, Ikom District, Cross North Forest Reserve, Ikom. High forest, fr. 8 June 1972, *J.A. Emwiogbon* in FHI 65823 (FHI, K!). **CAMEROON. Central Region:** Yaoundé, “Yaunde Station” 800 m, fl. 1890–1894, *Zenker & Staudt* 415 (B destroyed; BM, K!); **South Region:** Bipinde, Urwaldgebiet, fr. 1904, *Zenker* 2986 (BM!, BR!, K!, P!); **South West Region,** Kupe Muanenguba Division, Kupe Village, main trail towards Mount Kupe, forest near a valley, fr. 16 July 1996, *Etuge* 2798 (holotype K(K000109898)!; iso. BR(BR0000025613452V)!, MO, P, WAG(1966136)!, YA); *ibid*, main trail towards Mount Kupe, 800 m alt., fr. 16 July 1996, *Etuge* 2829 (K!, YA); Ndian Division, Rumpi Hills, ca. 6 km E of Dikome Balue on foot path to Ifanga Nalende, ca., 300 m E of junction with trail to Momboriba, in primary forest on clay loam with *Garcinia* and *Coelocaryon* spp., buds 10 Dec.1994, *Gereau, F. Namata, E. Jato, E. Sarabe*, 5639 (K!, MO, YA)

HABITAT & ECOLOGY. Submontane evergreen forest (where known); 800 – 940 m alt. The altitudes of two of the specimens cited above (from Cross River North and from Bipinde) are not given on the label so it is possible that they are from lower altitudes than the other specimens, where altitude is recorded. However, both locations include points that exceed 800 m altitude, so it is conceivable that they are consistent with the remaining specimens in this respect.

PHENOLOGY The initiation of flowering in December (dry season, *Gereau et al.* 5639) occurs at the same time as stem extension and is when new leaves are formed, while, when fruits are ripe (June and July, early wet season) the apical buds of the secondary stems appear dormant and there no new leaves are visible.

CONSERVATION STATUS. The relative frequency of occurrence of *Keetia nodulosa* is extremely low, indicating that even at its known locations it is extremely rare. At each location it is known from only a single collection, except at Mt Kupe where two collections are known. However, these two were collected on the same day, by the same team, on the same path up the mountain, and were 31 numbers apart. It may be that they were collected from the same plant, first in the morning on the way up, and then at the end of the day, on the way down. While at the Ikom location few collections of any plants have been made, at the Mt Kupe and Bipinde locations many thousands of herbarium specimens have been collected (e.g. *Cheek et al.* 2004), so if the species was not extremely rare, further records would be expected.

Keetia nodulosa is here provisionally assessed as Endangered (EN B2 ab(iii)) under the IUCN (2012) standard because five locations are known (see specimens examined above), each with observed or inferred imminent or actual threats of habitat clearance resulting from iron ore extraction infrastructure (Bipinde), quarrying and urbanisation (Yaoundé) and clearance for smallholder agriculture (Ikom, Rumpi Hills and Mt Kupe locations). *Keetia nodulosa* may already be extinct at the Yaoundé location due to the threats cited (*Murphy et al.* 2023). The area of occupation is assessed as 20 km², using the IUCN required 4 km² cell size. It is possible that the species also occurs in Gabon at Mt. Belinga (see notes below) but since the physical specimen, *Texier* 2164 has not been verified by the authors, only seen as an image (which shows some anomalous characters, see notes below), it is not included, taking the precautionary principle. *Keetia nodulosa* may yet be found in other locations within or outside the range documented here. However, the likelihood of this is not high, since tens of thousands of specimens have already been collected in surveys of suitable habitat in areas to

the north and south of, and also within its known range (Cheek *et al.* 1992; Cheek *et al.* 1996; Cable & Cheek 1998; Cheek *et al.* 1996; 2000; Maisels *et al.* 2000; Chapman & Chapman 2001; Harvey *et al.* 2004; Cheek *et al.* 2004; Cheek *et al.* 2006; Cheek *et al.* 2010; Harvey *et al.* 2010; Cheek *et al.* 2011; Murphy *et al.* 2023).

ETYMOLOGY. The species is named for the bacterial nodules conspicuous on the abaxial leaf surfaces of this species, in which it is currently unique in the genus, and in the tribe.

PHENOLOGY. Flowering, new leaves and stem extension in December (dry season); fruiting June and July (early wet season).

Notes

Keetia nodulosa is highly similar to *K. purseglovei*. The fruits, including the endocarps, which are so often useful in distinguishing species from each other in the genus, are more or less identical. It is not remarkable that material of the first species was included in the second.

In the protologue of *K. purseglovei*, five specimens from Cameroon are cited as paratypes, of which two are attributed here to *K. nodulosa* (see Specimens Examined above). A third Cameroonian paratype of *K. purseglovei*, Bates 1904 (Bitye, Ebolowa, BM! cited in error as 1940, but with a determination slip as *K. purseglovei* by Bridson) is a third, apparently undescribed species, differing from *K. nodulosa* in lacking bacterial nodules, in having suborbicular, strongly persistent stipules (vs subquadrate, caducous) and completely white, glossy primary stems (vs purple, streaked), and secondary stems completely glabrous in the flowering stage (vs sparse, patent, bristle hairs). This specimen also differs from *K. purseglovei* s.s. of Uganda, which has e.g. matt black primary stems, transversely elliptic mature stipules and much longer pedicels. Bates 1904 seems to represent yet another undescribed species. *Leeuwenberg* 5083 (60 km SW Eseka, BR image!, WAG image!) is a further paratype of *K. purseglovei*, also differing from *K. nodulosa* in lacking bacterial nodules. It appears to also differ from that species in lacking domatia, but this and other features needs to be confirmed by checking a physical specimen since even on the high quality images of BR, it is difficult to be certain. When this is possible, it may prove be conspecific with Bates 1904. Both specimens are lowland, c. 200 m alt. (vs 800 to 940 m alt. in *K. nodulosa*) and occur in southern Cameroon, between the Nyong and Ntem rivers. The remaining Cameroonian paratype, cited as Bates 1462 (BM) has not been found and neither has the remaining non Ugandan paratype of *K. purseglovei*, *Gossweiler* 9147 (BM, Zaire, Leopoldville Province) (Cheek pers. obs. Jan. 2024).

Searching gbif.org for *Keetia purseglovei* retrieves 41 records of which 31 have associated images, and which amount to 18 unique specimen records. Apart from those attributable to *Keetia purseglovei* sensu stricto (Uganda, two specimens studied, also likely three specimens from DRC subject to confirmation after physical examination) and *K. nodulosa* (four specimens cited in this paper), specimens are also from Cameroon (*Leeuwenberg* and Bates see attributions above), the Central African Republic (3), Republic of Congo (1), Angola (1) and Gabon (2). Inspection of associated images, where available, and where resolution permits, reveals that with one exception, none have the bacterial nodules of *K. nodulosa*. These specimens also show dissimilarities with *Keetia purseglovei*. It is possible that they also may represent further new species to science, potentially conspecific with Bates 1904 (see above).

Texier 2164 (BR, BRLU, G, LBV, MO, P, WAG) was collected at the Mt Belinga chain, 60 km NE of Makoukou. The images available on gbif.org of plants live in the field clearly show black bacterial nodules on the abaxial surfaces of the leaves

(<https://www.tropi...imageid=100597044>). Mt Belinga is known to host submontane forest,

the habitat of *K. nodulosa*. However, *Texier* 2164 has 1) densely hairy stems, atypical of *K. nodulosa*, which has sparse hairs on the stem at the flowering stage, 2) leaf blades with length:width ratio c. 4: 1 (vs 2 – 3: 1), which 3) lack an acumen, 4) inflorescences 1.5 times the petiole length (vs c.3 – 4 times). Taken together these differences suggest that *Texier* 2164 may be a second species of *Keetia* with bacterial nodules. Verification of a physical specimen is desirable to establish a firm identification.

Gereau et al. 5639 had previously been identified (not by Bridson) as “*Keetia cf. hispida* sensu lato (aff. *setosum* Hiern)”. This was no doubt due to the setose hairs. However, *Keetia hispida* s.l. has swollen, ant-inhabited primary stem nodes, larger leaves with domatia along the secondary nerves, and lacks the quadrangle stipule blades of *K. nodulosa*. *Gereau et al.* 5639 has only immature flower buds but is consistent with *K. nodulosa* in all features including the presence of bacterial nodules.

Variation within *Keetia nodulosa*. While the four fruiting specimens of *Keetia nodulosa* are relatively uniform morphologically, the sole specimen with open flowers, *Zenker & Staudt* 415 (“Yaunde-Station”) is slightly anomalous in that the leaves are longer (reaching 9 – 10 cm long vs <9 cm long). Only a small portion of one abaxial leaf is visible on the specimen, and this is insect-damaged, making unambiguous confirmation of the presence of bacterial nodules challenging. It is even possible that *Zenker & Staudt* 415 is taxonomically separable from the other specimens that comprise *Keetia nodulosa*

That bacterial nodules were not previously detected in specimens of *Keetia nodulosa*, two of which have been in herbaria for more than 100 years, is likely because there was no reason to expect them to be found. It was only the first author’s work identifying and describing other new species to science with bacterial nodules in the same location (Mt Kupe) and at about the same time (*Cheek et al.* 2008) that had raised awareness of this trait and facilitated its detection in the *Keetia* in 2004 (*Cheek et al.* 2004).

Discussion

Leaf bacterial nodulation

Since first reported (in *Pavetta*, Rubiaceae, Zimmerman 1902), knowledge of bacterial nodulation in leaves of flowering plants, occurring only in palaeotropical Primulaceae and Rubiaceae (but see notes on *Dioscorea* L. and *Stryrax* L. below), has been growing steadily. Reviews on the subject include Boodle (1923), Lersten and Horner (1976), Lemaire *et al.* (2011), Yang & Hu (2018), and Pinto-Carbó *et al.* (2018). ‘Bacterial leaf symbiosis’ is characterized as comprising endosymbiotic bacteria being organized in specialized leaf structures, usually known as nodules, or sometimes as galls, bacteriocecidia, or warts. These are visible macromorphological aspects of the symbiosis (Lemaire *et al.* 2011). The bacteria of the nodules are gram negative, rod or ellipsoid in shape, c. 2 micrometres long, and lack flagellae (Carlier *et al.* 2017). They are intercellular, and colonise the leaves through the stomata (Rubiaceae) or marginal teeth (Primulaceae) from the apical bud, from which inflorescences, flowers, and so eventually developing seeds, are also colonized. The symbiotic bacteria concerned have been placed in the genus *Burkholderia* s.l. (Pinto-Carbó *et al.* (2018). Bacterial colonization of leaves without the bacteria being organized into visible leaf structures also occurs, with the bacteria thinly scattered inside the leaf (endophytic) between the mesophyll cells (Verstraete *et al.* 2017). Such endophytic non BLN bacteria occur more widely in genera of Rubiaceae than do BLN and have been reported from

two non BLN genera of Coffeaeae (Verstraete *et al.* 2023) and five non BLN genera of the Vanguerieae, but were not found in *Keetia* species sampled (Verstraete *et al.* 2013). Transmission of bacteria between plants is known to be mainly vertical (Pinto-Carbó *et al.* 2018). However, in the Rubiaceae, though not in Primulaceae, there is evidence that horizontal transfer can also occur (Pinto-Carbó *et al.* 2018). It is speculated that this is effected by sap sucking insects moving from plant to plant, since the guts of some of these insects are known to be home also to *Burkholderia* bacteria. Lemaire *et al.* (2011) is a detailed recent study on the taxonomic occurrence of bacterial leaf nodulation in host plants. It is focused on the phylogenetic placement (genes 16S, rDNA, *recA*, and *gyrB*) of the bacteria (endosymbionts) of 54 plant species in four of the six known leaf nodulated plant genera (see below). This amounts to nearly 10% of all known nodulated plant species. The genera *Amblyanthus* A.DC and *Amblyanthopsis* Mez, both Primulaceae of Asia in which BLN are recorded, were not sampled. The study confirmed that free living, soil dwelling bacteria are basal in the clade *Burkholderia* s.l. and sister to the leaf nodulating species. In almost all cases of BLN symbiosis, there is a 1:1 relationship of a species of bacteria with a taxon of plant. Only one example is known of a plant species, *Psychotria kirkii* Hiern, which has been colonised twice, by different taxa of bacteria (Lemaire *et al.* 2011). The earliest branching subclade of *Burkholderia* s.l. to colonise plants is that inhabiting some Asian *Ardisia* Sw. species (Primulaceae, formerly Myrsinaceae, Larson *et al.* 2023). The next earliest branching subclade colonises some species of the genus *Sericanthe* Robbr. (Rubiaceae Coffeaeae, Cheek *et al.* 2018d), 11 to 12 of the 17 known species being considered to have nodules) and *Pavetta* (Rubiaceae Pavetteae De Block *et al.* 2015) of which 350/400 species are considered to have nodules). Another branch colonises several species of *Psychotria* L. (Rubiaceae, Psychotrieae, Lachenaud 2019) in which c. 80/1400 species are nodulated. The penultimate branches colonise mainly further species of the genus *Pavetta* but include colonisation of some other species of both *Sericanthe* and *Psychotria*. The final subclades colonise the majority of the *Psychotria* BLN species. Thus, the genera *Psychotria*, *Pavetta*, and *Sericanthe* have each been colonized more than once, independently, by bacteria likely either from the soil or from other plants. Therefore, there have been multiple horizontal transfers of bacteria to leaf nodulated plant species, and co speciation or evolution of endosymbionts with their host plants through vertical transfer has not been universal. Divergence estimates by Lemaire *et al.* (2011) point to a relatively recent origin of bacterial symbiosis in Rubiaceae, dating back to the Miocene (5 to 23 Mya).

Following strong support from genome analysis, the bacterial genus *Burkholderia* s.l. has been divided into several genera which largely correspond to different lifestyles or symbioses (Estrada de los Santos *et al.* (2018). *Burkholderia* s.s. are human and animal pathogens, while symbionts of the fungal phytopathogen *Rhizopus microsporus* are now classified as *Mycetohabitans*. *Mimosa* root nodulating bacteria are classified as *Trinickia*, and 'plant beneficial and environmental strains' (including the bacterial nodulating leaf symbionts discussed above) are now classified as *Paraburkholderia*, which genus includes also other N₂ fixing legume root symbionts. N₂ fixing legumes are also colonized by bacteria of the genus *Caballeronia*, but *Caballeronia* are also endophytic in the leaves of the non BLN genera *Empogona* Hook.f. and *Tricalysia* A.Rich. ex DC. of Coffeaeae (Verstraete *et al.* 2023). *Paraburkholderia* can also be symbionts of amoeba e.g. *Dictyostelium discoideum*, and of insect guts (Brock *et al.* 2020).

Bacterial leaf nodulation is also considered to occur in the tropical African monocot *Dioscorea sansibarensis* Pax (Dioscoreaceae), where folding of the leaf apices forms visible (pale green) pockets which allow development of bacterial colonies of *Orella dioscoreae*

(Alcaligenaceae, Burkholderiales, Carlier *et al.* 2017). Bacterial colonisation of marginal leaf glandular hairs has been observed in *Styrax camporum* Pohl of Brazil (Styracaceae, Machado *et al.* 2014), but the bacteria, which are both intra and intercellular, remain unidentified and nodules are not formed.

The endosymbiont bacteria of Rubiaceae have a small genome size and low coding capacity, both characteristic of reductive genome evolution. Genome sizes range from 2.4 Mb to 6.1Mb, well below the c.8 Mb average of free living *Burkholderia* s.l. species. Loss of functional capacity likely explains the failure of repeated efforts to cultivate endosymbiont bacteria (Pinto-Carbó *et al.* 2018). Equally, cultivated plants which lack their endosymbionts grow poorly and eventually die (Verstraete *et al.* 2017).

Although the genome of endosymbionts is reduced, synthesis gene clusters have been detected in those of all *Psychotria* and *Pavetta* species investigated so far (Pinto-Carbó *et al.* 2018). Evidence that the novel C₇N aminocyclitol kirkamide is synthesized by the symbiont bacteria in *Psychotria kirkii* is that while it is detected in leaves of plants with the endosymbiont, it is not in aposymbiotic plants (lacking the endosymbiont). The compound is toxic to arthropods and insects, suggesting a role in protecting the host against herbivory (Sieber *et al.* 2015). A related compound, streptol glucoside is also found in the nodulated leaves of the same species. It displays potent herbicidal activity and may have allelopathic properties (Pinto-Carbó *et al.* 2018). Presence of such bacterial endosymbionts may thus be advantageous for the hosts and confer an evolutionary advantage over plants which lack such endosymbionts. We can hypothesise that because species with bacterial leaf nodules contain many more bacteria than non BLN species, the quantity of advantageous compounds produced by the bacteria might be higher, increasing the evolutionary advantage further.

The bacterial nodules in Rubiaceae vary in form from genus to genus, and also within genera. In *Psychotria* they usually black, raised and conspicuous to the naked eye on the abaxial leaf surface, scattered uniformly over the blade, the shape, size and density of the nodules helping to separate one species from another. In a minority of species the nodules are linear and positioned next to the midrib only (Lachenaud 2019; Cheek *et al.* 2008). In contrast, in *Pavetta*, the nodules are usually most conspicuous on the adaxial surface, also black but in other species green and inconspicuous unless viewed in transmitted light. Frequently they occur as thickenings at the junction of the tertiary nerves (Manning 1996). In *Sericanthe*, the nodules are often inconspicuous unless viewed in transmitted light, and often linear and arranged along the midrib (e.g. Sonké *et al.* 2012). The regular pattern and spacing of the nodules through the leaf identifies them as such and differentiates BLN from e.g. epidermal fungal colonies which are more localized to only part of a leaf.

In herbarium specimens of *Keetia nodulosa* the nodules have similarities with those commonly seen in *Pavetta* see above. They are black, conspicuous, slightly raised, and often at nerve junctions. However, they differ from most *Pavetta* in being conspicuous only abaxially, as in the BLN of *Psychotria*.

The discovery of bacterial nodules in a further tribe and genus of Rubiaceae was unexpected. A survey of the occurrence of endosymbiotic bacteria specifically in the Vanguerieae found that they only occur in five genera, in none of which are nodules formed, and none of which were *Keetia* (Verstraete *et al.* 2017).

Further work is needed to identify the species of bacterium that produces the nodules in *Keetia*. This can be done by genomic studies of dried leaf material (Danneels & Carlier

2023). The symbiont is almost certain to be a *Paraburkholderia*, given that all other leaf nodule forming endosymbionts of Rubiaceae belong to this genus, and that the non BLN endophytic bacteria recorded in Vanguerieae are also this genus (Verstraete *et al.* 2017). It will be especially interesting to find out in which subclade of *Paraburkholderia* it falls, and so to deduce the source and date of this colonization. It can be speculated that the colonization event is recent, since this is the only known nodule-forming species in a genus of 40 species. If the event was as old as in the other nodulated genera of Rubiaceae (see above), one might expect that a much higher number, and proportion of the species, would have been found to have been nodulated, as in those other three genera. We speculate that the event may have occurred in the vicinity of the Cross-Sanaga Interval (Cheek *et al.* 2001) which has the highest species and generic diversity per degree square in tropical Africa (Barthlott *et al.* 1996; Dagallier *et al.* 2020). Here, three of the five locations of *Keetia nodulosa* occur, two others being nearby). All three of the other Rubiaceae genera with bacterial nodules have centres of species diversity in the Cross-Sanaga Interval (Lachenaud 2019; Manning 1986; Sonke *et al.* 2012) from which horizontal transfer to *Keetia* mediated by sap-sucking insects may have occurred.

The discovery reported in this paper of bacterial leaf nodulation in a genus and tribe previously unknown to have this characteristic, is the first since the report 60 years ago by Petit (1962) of nodulation in some species he attributed to *Tricalysia* which are now placed in *Sericanthe*. It is conceivable that bacterial leaf nodulation remains to be found in other genera in which it is previously currently unknown.

Submontane forest species in the western half of Cameroon

Additional rare, threatened species of submontane forest found with *Keetia nodulosa* at Mt Kupe, Rumpi Hills, or elsewhere within the range of the species are *Coffea montekupensis* Stoffel. (Rubiaceae, Stoffelen *et al.* 1997), *Psychotria hardyi* O.Lachenaud (Rubiaceae, Lachenaud 2019), *Memecylon kupeanum* R.D.Stone *et al.* (Melastomataceae, Stone *et al.* 2008), *Sabicea bullata* Zemagho *et al.* (Rubiaceae, Zemagho *et al.* 2014), *Impatiens frithii* Cheek (Balsaminaceae, Cheek & Csiba 2002), *Microcos magnifica* Cheek (Cheek 2017) and *Microcos rumpi* Cheek (Cheek *et al.* 2023a) both Malvaceae s.l./Grewiaceae, *Cola etugei* Cheek (Malvaceae s.l./Sterculiaceae, Cheek *et al.* 2020b), *Psychotria spp.* (Rubiaceae, Cheek *et al.* 2008), *Deinbollia oreophila* Cheek (Sapindaceae, Cheek & Etuge 2009), *Kupea martinetegei* Cheek (Triuridaceae, Cheek *et al.* 2003), and *Vepris zapfackii* Cheek (Rutaceae, Cheek & Onana 2021). In several cases the species were initially considered point endemics but were shown after further surveys, to be more widespread in the surviving cloud forests of western Cameroon. However, in other cases despite additional surveys, species have remained point endemics e.g. *Brachystephanus kupeensis* I.Darbysh. (Acanthaceae, Champluvier & Darbyshire 2009). The high level of endemism in these submontane forests (extending to Bioko) contributes to the high species and generic diversity levels reported in the Cross Sanaga Interval mentioned above. There is no doubt that additional species remain to be described from these forests, although further survey work is hampered by the secession struggle in the two anglophone Regions, South West and North West that began in December 2016 and has taken thousands of lives and displaced half a million people (https://en.wikipedia.org/wiki/Anglophone_Crisis, accessed Feb. 2024). South West Region contains the majority of the surviving submontane forest in western Cameroon, indeed in the whole of the Gulf of Guinea.

Keetia nodulosa is one of many new species to science that came to light partly or entirely as a result of surveys for conservation prioritization in Cameroon. Cameroon has the highest

number of globally extinct plant species of all countries in continental tropical Africa (Humphreys *et al.* 2019). The extinction of species such as *Oxygyne triandra* Schltr. (Thismiaceae, Cheek *et al.* 2018b) and *Afrothisia pachyantha* Schltr. (Afrothismiaceae, Cheek & Williams 1999; Cheek *et al.* 2019; Cheek *et al.* 2023d) and at least two species of the African genus *Inversodicraea* (Cheek *et al.* 2017), are well known examples, recently joined by species such as *Vepris bali* Cheek (Rutaceae, Cheek *et al.* 2018c), *Vepris montisbambutensis* Onana (Onana & Chevillotte 2015) and *Ardisia schlechteri* Gilg (Murphy *et al.* 2023). However, another 127 potentially globally extinct Cameroon species have recently been documented (Murphy *et al.* 2023: 18 – 22).

It is critical now to detect, delimit and formally name species such as *Keetia nodulosa* as new to science, since until they are scientifically recognised, they are essentially invisible to science, and only when they have a scientific name can their inclusion on the IUCN Red List be facilitated (Cheek *et al.* 2020a). Most (77%) species named as new to science in 2023 are already threatened with extinction (Brown *et al.* 2023). Many new species to science have evaded detection until today because they are in genera that are long overdue full taxonomic revision as was the case with *Keetia nodulosa*, or because they have minute ranges which have remained unsurveyed until recently.

If further global extinction of plant species is to be avoided, effective conservation prioritization is crucial, backed up by investment in protection of habitat, ideally through reinforcement and support for local communities who often effectively own and manage the areas concerned. Important Plant Areas (IPAs) programmes, often known in the tropics as TIPAs (Darbyshire *et al.* 2017; Couch *et al.* 2019; Darbyshire *et al.* 2023; Murphy *et al.* 2023) offer the means to prioritize areas for conservation based on the inclusion of highly threatened plant species, among other criteria. Such measures are vital if further species extinctions are to be avoided of rare, highly threatened species such as *Keetia nodulosa*.

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