

Evaluation Of Medicinal Uses, Phytochemistry And Pharmacological Properties Of *Dioscorea Sylvatica* (Dioscoreaceae)

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Abstract: *Dioscorea sylvatica* Eckl. is a twining herb widely used as traditional medicine in southern Africa. The current study critically reviewed the medicinal uses, phytochemistry and pharmacological properties of *D. sylvatica*. A systematic review of the literature studies was carried out to document the medicinal uses, phytochemistry and pharmacological properties of *D. sylvatica*. The results of the current study are based on literature survey conducted using various search engines such as Web of Science, Elsevier, Pubmed, Google scholar, Springer, Science Direct, Scopus, Taylor and Francis, and pre-electronic sources such as books, book chapters, scientific journals and other grey literature obtained from the University library. This study revealed that *D. sylvatica* is used mainly as blood purifier, charm, rituals and ethnoveterinary medicine, traditional medicine for gastro-intestinal problems, gonorrhoea, respiratory infections, rheumatism, skin infections, sores, wounds and cuts. Pharmacological research identified 2-carboxyarabinitol, alkaloids, anthocyanins, bibenzyl, calcium oxalate crystals, diarylnonanoids, flavonoids, pyrogallol, sapogenin, saponins, shikimic acid, tannins and terpenoids. The crude extracts of *D. sylvatica* and phytochemical compounds isolated from the species exhibited antibacterial, antimycobacterial, antifungal, anti-inflammatory, antioxidant and antibiotic-potentiating activities. *Dioscorea sylvatica* should be subjected to detailed phytochemical, pharmacological and toxicological evaluations aimed at correlating its medicinal uses with its phytochemistry and pharmacological activities.

Keywords: *Dioscorea sylvatica*, Dioscoreaceae, ethnopharmacology, herbal medicine, indigenous knowledge, southern Africa

1 INTRODUCTION

Dioscorea sylvatica Eckl. (Fig. 1) is a twining herb belonging to the Dioscoreaceae family. The genus *Dioscorea* Plum ex L. is a genus of over 600 tuberous herbaceous climbing species which naturally occurs throughout the tropical and warm temperate regions of the world [1,2]. The genus name *Dioscorea* is in honour of Pedanius Dioscorides (40 – 90 AD), a Greek author, botanist, pharmacologist and physician [3]. The species name *sylvatica* means “growing in woods” or “forest-loving” [4]. The synonyms associated with the name *D. sylvatica* include *D. brevipes* Burt Davy, *D. hederifolia* Griseb., *D. marlothii* R. Knuth, *D. montana* (Burch.) Spreng var. *glauca* R. Knuth, *D. montana* (Burch.) Spreng var. *lobata* Weim., *D. montana* (Burch.) Spreng var. *sagittata* Suess., *D. rehmannii* Baker, *D. sylvatica* ssp. *lydenbergensis* Blunden, Hardman & F.J. Hind, *D. sylvatica* (Kunth) Eckl. var. *brevipes* (Burt Davy) Burkill, *D. sylvatica* (Kunth) Eckl. var. *sylvatica*, *Testudinaria glaucescens* Hügel, *T. multiflora* Marloth, *T. paniculata* Dummer, *T. rehmannii* (Baker) G.D. Rowley, *T. sylvatica* Kunth, *T. sylvatica* var. *brevipes* (Burt Davy) G.D. Rowley, *T. sylvatica* var. *lydenbergensis* (Blunden, Hardman & F.J. Hind) G.D. Rowley, *T. sylvatica* var. *multiflora* (Marloth) G.D. Rowley, *T. sylvatica* var. *paniculata* (Dummer) G.D. Rowley and *T. sylvatica* var. *rehmannii* (Baker) G.D. Rowley [5,6]. *Dioscorea sylvatica* is closely related to *D. elephantipes* (L'Hér.) Engl. as the two species are characterised by abundance of phytochemical compounds shikimic acid and pyrogallol [7]. *Dioscorea sylvatica* has alternate, heart-shaped and often tapering to a long point at the apex [8] (Figure 1). The underground tubers of *D. sylvatica* are characteristically dark brown, corky and flattened, with reticulate markings [9]. *Dioscorea sylvatica* has been recorded in Eswatini, Mozambique, South Africa, Zambia and Zimbabwe in bushland, rocky outcrops, dunes, woodland, coastal regions and forest margins at an altitude ranging from 125 m to 1650 m above sea level [6,10,11].



Fig. 1: *Dioscorea sylvatica*: A: branch showing leaves and flowers and B: flowers (photos: BT Wursten)

Dioscorea sylvatica is a valuable medicinal plant in southern Africa and an overview of its botanical description, active ingredients, pharmacological effects and distribution are outlined in the monograph “medicinal plants of South Africa” [12]. *Dioscorea sylvatica* is cultivated in home gardens as a source of traditional medicines in the Limpopo province, South Africa [13,14]. Several *Dioscorea* species contain a spirostane steroidal sapogenin known as diosgenin, used to manufacture cortisone and other steroid hormones [15] with *D. zingiberensis* C.H. Wright (Chinese yam) being the commercial source of the phytochemical compound [16]. Moreover, a patent highlighting the production of a phytochemical compound shikimic acid from aerial parts of *D. sylvatica* as well as other *Dioscorea* species such as *D. elephantipes*, *D. sylvatica*, *D. villosa* L., *D. communis* (L.) Caddick & Wilkin, *D. caucasia* Lipsky, *D. nipponica* Makino, *D. deltoidea* Wall. Ex Griseb., *D. tokoro* Makino ex Miyabe, *D. prussi* Pax, *D. alata* L., *D. altissima* L., *D. sansibarensis* Pax,

D. bulbifera L., *D. antaly* Jum. & H. Perrier, *D. prae-hensis* Benth. and *D. minutiflora* Mart. ex Griseb. [17]. A tremendous population decline of *D. sylvatica* was recorded in South Africa from 1955-1960 due to indiscriminate commercial harvesting for diosgenin [18-23]. Research by Raimondo et al. [22] showed that *D. sylvatica* is categorized as Vulnerable A2cd under the IUCN Red List Categories and Criteria version 3.1 (<http://www.iucnredlist.org>). *Dioscorea sylvatica* is sold in informal herbal medicine markets in Eswatini and South Africa [24-38]. Wiersum et al. [39] recommended *D. sylvatica* for cultivation in home gardens to serve as a tool for combined biodiversity conservation and poverty alleviation in South Africa. It is therefore, within this context that this study was undertaken aimed at reviewing the medicinal uses, phytochemistry and pharmacological properties of *D. sylvatica*.

2. MATERIALS AND METHODS

Several electronic databases were searched which included Web of Science, Elsevier, Pubmed, Google scholar, Springer, Science Direct, Scopus, Taylor and Francis. Additional information was obtained from pre-electronic sources such as books, book chapters, scientific journals and other grey literature obtained from the University library. The relevant terms *Dioscorea sylvatica* was paired with keywords such as "medicinal uses of *Dioscorea sylvatica*", "phytochemicals of *Dioscorea sylvatica*", "biological activities of *Dioscorea sylvatica*", "pharmacological properties of *Dioscorea sylvatica*", "ethnobotany of *Dioscorea sylvatica*", and various other synonyms and common names of the plant species. The ultimate goal of this search was to explore articles that investigated the medicinal uses, phytochemical and pharmacological properties of *D. sylvatica*. A total of 40 articles published between 1954 and 2021 matched the inclusion criteria and were included in this review (Fig. 2).

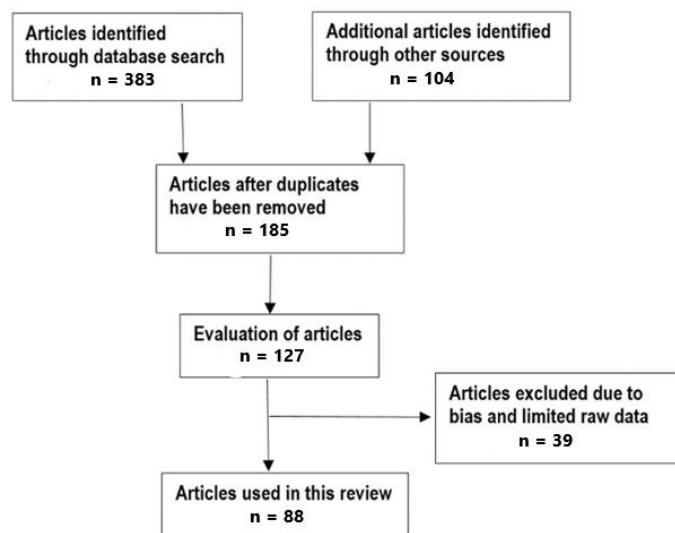


Fig. 2. Flow chart showing the number of research publications used in this study

3. RESULTS AND DISCUSSION

3.1 Medicinal uses of *Dioscorea sylvatica*

The roots and tubers of *D. sylvatica* are mainly used as blood purifier, charm, rituals and ethnoveterinary medicine,

traditional medicine for gastro-intestinal problems, gonorrhoea, respiratory infections, rheumatism, skin infections, sores, wounds and cuts (Table 1; Fig. 3). Other medicinal applications of the roots and tubers of *D. sylvatica* supported by at least two literature records epilepsy [40,41], facilitation of labour [41,42], parasite repellent [43,44] and swellings [43,44].

TABLE 1
MEDICINAL USES OF DIOSCOREA SYLVATICA

Medicinal uses	Plant part	Country	References
Blood pressure	Tuber	South Africa	45
Blood purifier	Roots and tubers	Eswatini and South Africa	36,41,43,44, 46-49
Body pains	Tuber	South Africa	38
Charm and ritual (good luck, protective and ritual emetics)	Roots	South Africa	30,36,40,47, 48,50,51
Contraceptive	Tubers	South Africa	42
Epilepsy	Roots and tubers	South Africa	40,41
Facilitate labour	Tubers	South Africa	41,42
Fibroids	Tuber	South Africa	52
Foot disorder	Tubers	South Africa	38
Gastro-intestinal problems (abdominal cramps and stomach ache)	Tuber	South Africa and Zimbabwe	41,53
Gonorrhoea	Roots and tuber	South Africa	13,14,54
Gonorrhoea	Tuber mixed with roots of Aloe marlothii A. Berger	South Africa	55,56
Heart problems	Tuber	South Africa	30
Hysteria	Tuber	South Africa	41
Infertility	Tuber	South Africa	41
Inflammation	Roots and tuber	South Africa	57
Leg pains	Tuber	South Africa	36
Nervous spasms	Tuber	South Africa	42
Parasite repellent	Roots	South Africa	43,44
Respiratory infections (aphonia, asthma, bronchiectasis, chest complaints and tuberculosis)	Roots and tubers	Eswatini and South Africa	11,36,41,43, 46-49,58-67
Rheumatism	Tuber	South Africa and Zimbabwe	52,53,58,68-70
Skin infections (abscess, pimples, rashes and skin complexion)	Tubers and whole plant	Eswatini, South Africa and Zimbabwe	42,47,48,53, 58,69-74
Sores, wounds and cuts	Leaves and tubers	South Africa	8,29,31,40-42,52,75-78
Strengthen muscles	Tuber	South Africa	30
Stroke	Tuber	South Africa	30
Swellings	Tubers	South Africa	45,47,48
Vomiting	Roots	South Africa	40
Ethnoveterinary medicine (sores and wounds, udder mastitis in cows and uterine problems in cows)	Tuber	South Africa	31,36,42,43, 49,58,77,79-82

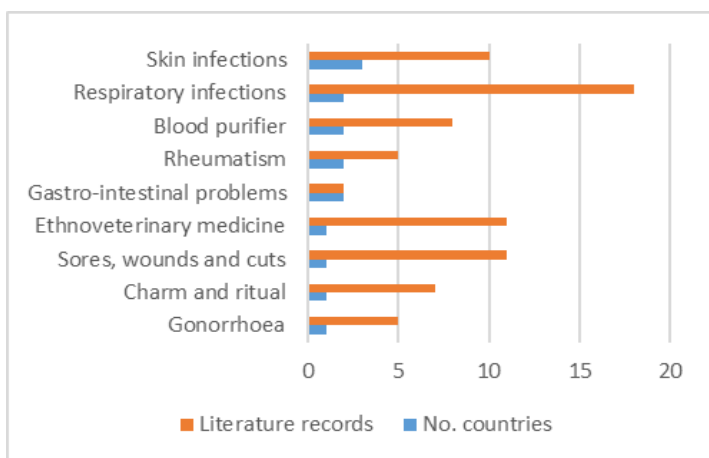


Fig. 3: Medicinal uses of *Dioscorea sylvatica* in southern Africa

3.2 Phytochemistry of *Dioscorea sylvatica*

A variety of chemical compounds have been isolated and identified from *D. sylvatica* (Table 2). These phytochemical compounds identified from the aerial parts, tubers and whole plant parts of *D. sylvatica* include 2-carboxyarabinitol, alkaloids, anthocyanins, bibenzyl, calcium oxalate crystals, diarylnonanoids, flavonoids, pyrogallol, sapogenin, saponins, shikimic acid, tannins and terpenoids (Table 2).

TABLE 2
PHYTOCHEMICAL COMPOUNDS ISOLATED FROM
DIOSCOREA SYLVATICA

Phytochemical compound	Value	Plant part	Reference
2-carboxyarabinitol (nmol/g)	99.0	Leaves	83
5-(2-hydroxyphenethyl)-2,3-dimethoxyphenol	-	Tubers	84,85
Alkaloids (%)	9.9	Whole plant	42
Anthroquinones	-	Whole plant	42
Bibenzyl	-	Tubers	85
Calcium oxalate crystals	-	Tubers	69
(E)-32-((3-(3-hydroxy-4-methoxyphenyl)isoferuloyl)oxy)dotriacontanoic acid	-	Tubers	84,85
Flavonoids	-	Whole plant	42,73
Pyrogallol	-	Aerial parts	7
Sapogenin	-	Tubers	86,87
Saponins (%)	9.0	Whole plant	42,73
Shikimic acid	-	Aerial parts	7
Steroids	-	Whole plant	42
Tannins	-	Whole plant	42,73
Terpenoids	-	Whole plant	42,73

3.3 Pharmacological properties of *Dioscorea sylvatica*

The following biological activities have been reported from the roots, tubers, tuber bark and whole plant parts of *D. sylvatica* and phytochemical compounds isolated from the species: anti-acetylcholinesterase [70], antibacterial [42,58,70,73,77], antimycobacterial [58,88], antifungal [42,58,70,73,88], anti-inflammatory [42], antioxidant [42,70] and antibiotic-potentiating [84,85] activities.

3.3.1 Anti-acetylcholinesterase activities

Cogne [70] evaluated the anti-acetylcholinesterase activities of petrol ether, water, methanol, dichloromethane, ethyl acetate and butanol extracts of *D. sylvatica* tuber using a rapid TLC

bioautographic method for the detection of acetylcholinesterase. The petrol ether, dichloromethane and ethyl acetate extracts exhibited moderate activities [70].

3.3.2 Antibacterial activities

Kelmanson et al. [77] evaluated the antibacterial activities of methanol, ethyl acetate and water extracts of *D. sylvatica* roots, tubers and tuber bark against *Bacillus subtilis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Micrococcus luteus*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Staphylococcus epidermidis* using the disc diffusion assay and twofold serial dilution with neomycin (2.0 µg) as a positive control. The ethyl acetate and methanol extracts exhibited activities against *Bacillus subtilis*, *Escherichia coli*, *Micrococcus luteus*, *Staphylococcus aureus* and *Staphylococcus epidermidis* with minimal inhibitory concentration (MIC) values ranging from 2.0 mg/ml to 4.0 mg/ml [77]. Cogne [70] evaluated the antibacterial activities of petrol ether, water, methanol, dichloromethane, ethyl acetate and butanol extracts of *D. sylvatica* tuber against *Bacillus subtilis* using a rapid TLC bioautographic method. The petrol ether, dichloromethane and ethyl acetate extracts exhibited moderate activities [70]. Seaman [58] evaluated the antibacterial activities of acetone and methanol extracts of *D. sylvatica* tubers against *Staphylococcus aureus*, *Enterococcus faecalis*, *Bacillus cereus*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Serratia odorifera* and *Moraxella catarrhalis* using broth microdilution method with neomycin and ciprofloxacin as positive controls. The extracts showed activities against the tested pathogens with MIC values ranging from 0.5 mg/ml to 8.0 mg/ml [58]. Xaba [42] and Xaba and Buwa [73] evaluated the antibacterial activities of methanol, ethanol, acetone and aqueous extracts of *D. sylvatica* whole plant against *Pseudomonas aeruginosa*, *Bacillus pumilus*, *Staphylococcus aureus*, *Escherichia coli*, and *Klebsiella pneumoniae* using microplate dilution assay. The extracts exhibited activities against tested pathogens with MIC values ranging between 3.2 mg/ml and 12.5 mg/ml [42,73].

3.3.3 Antimycobacterial activities

Seaman [58] evaluated the antimycobacterial activities of acetone and aqueous leaf extracts of *N. ivifolia* against *Mycobacterium smegmatis* and *Mycobacterium aurum* using broth microdilution technique and *Mycobacterium tuberculosis* using BACTEC susceptibility testing with rifampicin and ciprofloxacin as positive controls. The extracts exhibited activities with MIC values ranging from 0.3 mg/ml to 8.0 mg/ml [58]. Lehasa et al. [88] evaluated the antimycobacterial activities of aqueous and ethanol extracts of *D. sylvatica* tubers against *Mycobacterium tuberculosis*. The extract exhibited activities against tested pathogen with MIC values ranging between 0.2 mg/ml and 0.8 mg/ml [88].

3.3.4 Antifungal activities

Cogne [70] evaluated the antifungal activities of petrol ether, water, methanol, dichloromethane, ethyl acetate and butanol extracts of *D. sylvatica* tuber against *Cladosporium cucumerinum* and *Candida albicans* using a rapid TLC bioautographic method. The petrol ether and dichloromethane extracts exhibited weak activities against *Cladosporium cucumerinum* [70]. Seaman [58] evaluated the antifungal activities of methanol and acetone extracts of *D. sylvatica* tubers against *Candida albicans* using the broth microdilution

method with nystatin as a positive control. The methanol and acetone extracts exhibited activities with MIC value of 4.0 mg/ml [58]. Xaba [42] and Xaba and Buwa [73] evaluated the antifungal activities of methanol, ethanol, acetone, and aqueous extracts of *D. sylvatica* whole plant against *Candida albicans* and *Trichophyton mucoides* using microplate dilution assay. The extracts exhibited activities against tested pathogens with an MIC values ranging from 0.4 mg/ml to 3.1 mg/ml [42,73]. Lehasa et al. [88] evaluated the antifungal activities of aqueous and ethanol extracts of *D. sylvatica* tubers against *Candida albicans* and *Trichophyton mucoides* using microplate dilution assay. The extract exhibited activities against tested pathogen with MIC values ranging between 0.05 mg/ml and 0.7 mg/ml [88].

3.3.5 Anti-inflammatory activities

Xaba [42] evaluated the anti-inflammatory activities of aqueous, acetone, ethanol and methanol extracts of *D. sylvatica* whole plant using the 5-lipoxygenase enzyme (Cayman) assay with nordihydroguaiaretic acid as a positive control. The extracts exhibited activities with percentage inhibition of enzyme of >85.0% and half maximal inhibitory concentration (IC₅₀) values ranging from 0.2 µg/ml and 0.7 µg/ml, which was comparable to IC₅₀ value of 0.6 µg/ml exhibited by the positive control [42].

3.3.6 Antioxidant activities

Cogne [70] evaluated the antioxidant activities of petrol ether, water, methanol, dichloromethane, ethyl acetate and butanol extracts of *D. sylvatica* tuber using 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical scavenging assay. The ethyl acetate extract exhibited moderate activities [70]. Xaba [42] evaluated the antioxidant activities of acetone, aqueous, ethanol, and methanol extracts of *D. sylvatica* whole plant using DPPH free radical scavenging and the phosphomolybdate assays with ascorbic acid and gallic acid as positive controls. The extracts exhibited activities with IC₅₀ values ranging from 0.1 µg/ml to 0.4 µg/ml against DPPH and IC₅₀ values of 0.04 µg/ml to 0.8 µg/ml against phosphomolybdate assay [42].

3.3.7 Antibiotic-potentiating activities

Sibandze et al. [84] and Sibandze et al. [85] evaluated the antibiotic-potentiating activities of the compounds (E)-32-((3-(3-hydroxy-4-methoxyphenyl)isoferuloyloxy)dotriacontanoic acid and 5-(2-hydroxyphenethyl)-2,3-dimethoxyphenol isolated from *D. sylvatica* tubers using two multi-drug resistant *Staphylococcus aureus* strains SA-1199B and XU212 with reserpine as positive control. The compounds exhibited potent antibiotic-potentiating activities [84,85].

4. CONCLUSION

The present review summarizes the ethnomedicinal uses, phytochemistry and pharmacological properties of *D. sylvatica*. The historical traditional usage of *D. sylvatica* as herbal medicine in southern Africa calls for detailed phytochemical and pharmacological studies aimed at correlating its documented ethnomedicinal uses with the phytochemical and pharmacological properties of the species. There is need for clinical and toxicological evaluations of both crude extracts and phytochemical compounds associated with *D. sylvatica*.

Dioscorea sylvatica has been traditionally used as herbal medicine throughout its distributional range in sub-Saharan Africa, used as blood purifier, charm, rituals and ethnoveterinary medicine, traditional medicine for gastrointestinal problems, gonorrhoea, respiratory infections, rheumatism, skin infections, sores, wounds and cuts. Recent research on *D. sylvatica* focused primarily on evaluating the anti-acetylcholinesterase, antibacterial, antimycobacterial, antifungal, anti-inflammatory, antioxidant and antibiotic-potentiating activities. The 2-carboxyarabinitol, alkaloids, anthocyanins, bibenzyl, calcium oxalate crystals, diarylnonanoids, flavonoids, pyrogallol, sapogenin, saponins, shikimic acid, tannins and terpenoids appear to be the major plant derivatives and have been demonstrated to be the main active ingredients in aerial parts, tubers and whole plant parts of *D. sylvatica*. Although contemporary research involving *D. sylvatica* is promising, it is too preliminary and sometimes too general to be used to explain and support some of the ethnomedicinal uses. In addition to this, some of the pharmacological activities assessed so far, were routine screenings using standard procedures lacking molecular mechanisms of the pharmacological effects of *D. sylvatica*. There is not yet enough systematic data regarding the pharmacokinetics and clinical research of *D. sylvatica* products and compounds. There are also very few to nil experimental animal studies, randomized clinical trials and target-organ toxicity studies involving *D. sylvatica* and its derivatives that have been carried out so far. Therefore, there is not sufficient evidence to interpret the documented ethnomedicinal uses linking them to specific chemical mechanisms associated with some of the documented biological activities of the species. Therefore, future studies should identify the bioactive components, details of the molecular modes or mechanisms of action, pharmacokinetics and physiological pathways for specific bioactives of *D. sylvatica*. Further research should comprise of the models which would more precisely refer to the current knowledge about pathophysiology, extensive phytochemical, pharmacological, preclinical and clinical research on well documented and established ethnomedicinal uses of the species. This future research should also include the identification of any side effects and/or toxicity, aspects of quality control to ensure safety, quality and efficacy of the *D. sylvatica* products.

CONFLICTS OF INTEREST

No conflict of interest is associated with this work.

REFERENCES

- [1] Govaerts R, Wilkin P and Saunders RMK. World checklist of Dioscoreales: Yams and their allies. Kew Publishing, Kew, Richmond, United Kingdom; 2007.
- [2] Christenhusz MJM and Byng JW. The number of known plants species in the world and its annual increase. *Phytotaxa*. 2016; 261: 201–217.
- [3] Mothogoane MS. *Dioscorea dregeana* (Kunth) T. Durand & Schinz; 2013. Available at: <http://pza.sanbi.org/dioscorea-dregeana>, accessed on 15 February 2021.
- [4] Glen HF. Sappi: What's in a name: The meanings of the botanical names of trees. Jacana Media Publication, Johannesburg, South Africa; 2004.
- [5] Von Teichman und Logischen I, Van der Schijff HP and Robbertse PJ. The genus *Dioscorea* L. in South Africa.

- Boissiera. 1975; 24: 215–224.
- [6] Wilkin P. Dioscoreaceae. In: Timberlake JR and Martins ES (Eds.), *Flora Zambesiaca*. Royal Botanic Gardens, Kew, Richmond, United Kingdom; 2009, pp. 109–140.
- [7] Price EJ, Wilkin P, Sarasan V and Fraser PD. Metabolite profiling of *Dioscorea* (yam) species reveals underutilised biodiversity and renewable sources for high-value compounds. *Scientific Reports*. 2016; 6: 29136.
- [8] Crouch N, Symmonds R, Spring W and Diederichs N. Fact sheets for growing popular medicinal plant species. In: Diederichs N (Ed.), *Commercialising medicinal plants: A southern African guide*. Sun Press, Stellenbosch, South Africa; 2006, pp. 97-142.
- [9] Pooley E. A field guide to wild flowers KwaZulu Natal region and the Eastern Region. Natal Flora Publications Trust, Durban, South Africa; 1998.
- [10] Germishuizen G and Meyer NL. *Plants of southern Africa: An annotated checklist*. Pretoria: *Strelitzia* 14, National Botanical Institute; 2003.
- [11] Pooley E. A field guide to wild flowers KwaZulu–Natal and the Eastern Regions. Natal Flora Publications Trust, Durban; 2005.
- [12] Van Wyk B-E, Van Oudtshoorn B and Gericke N. *Medicinal plants of South Africa*. Briza Publications, Pretoria; 2013.
- [13] Semenya SS and Potgieter MJ. Medicinal plants cultivated in Bapedi traditional healers' homegardens, Limpopo province, South Africa. *African Journal of Traditional, Complementary and Alternative Medicines*. 2014; 11: 126-132.
- [14] Semenya SS and Mokgoebo MJ. The utilization and conservation of indigenous wild plant resources in the Limpopo province, South Africa. In: Rhodes ER and Naser H (Eds.), *Natural resources management and biological sciences*. IntechOpen, London; 2020, pp. 112–167.
- [15] Raju J and Rao CV. Diosgenin, a steroid saponin constituent of yams and fenugreek: Emerging evidence for applications in medicine. In: Rasooli I (Ed.), *Bioactive compounds in phytomedicine*. IntechOpen, London; 2012, pp. 125–142.
- [16] Yi T, Fan LL, Chen H-L, Zhu GY, Suen HM, Tang Y-N, Zhu L, Chu C, Zhao Z-Z and Chen H-B. Comparative analysis of diosgenin in *Dioscorea* species and related medicinal plants by UPLC-DAD-MS. *BMC Biochemistry*. 2014; 15: 9.
- [17] Price EJ and Fraser PD. *Phytochemical recovery from plants*. World Intellectual Property Organization. WO 2017/118760 A1, issued July 13, 2017.
- [18] Codd LE. Drugs from wild yams. *African Wildlife*. 1960; 14: 215–225.
- [19] Archibald EEA. The genus *Dioscorea* in the Cape province west of East London. *Journal of South African Botany*. 1967; 33: 1-46.
- [20] Cunningham AB. An investigation of the herbal medicine trade in Natal/KwaZulu. Investigational Report No. 29. Institute of Natural Resources, Pietermaritzburg; 1988.
- [21] Von Ahlefeldt D, Crouch NR, Nichols G, Symmonds R, McKean S, Sibiyi H and Cele MP. Medicinal plants traded on South Africa's eastern seaboard. Porcupine Press, Durban; 2003.
- [22] Raimondo D, Von Staden L, Foden W, Victor JE, Helme NA, Turner RC, Kamundi DA and Manyama PA. Red List of South African plants. *Strelitzia* 25. South African National Biodiversity Institute, Pretoria, 2009.
- [23] Hills R, Muasya AM, Maurin O and Wilkin P. A threatened new species of *Dioscorea* from KwaZulu-Natal, South Africa, *Dioscorea hurteri* (Dioscoreaceae). *Kew Bulletin*. 2018; 73: 14.
- [24] Cunningham AB. *African medicinal plants: setting priorities at the interface between conservation and primary health care*. People and Plants working paper 1, UNESCO, Paris; 1993.
- [25] Mander M. Marketing of indigenous medicinal plants in South Africa: A case study in KwaZulu-Natal. Food and Agriculture Organization, Rome; 1998.
- [26] Williams VL, Balkwill K and Witkowski ETF. A lexicon of plants traded in the Witwatersrand umuthi shops, South Africa. *Bothalia*. 2001; 31: 71-98.
- [27] Dold AP and Cocks ML. The trade in medicinal plants in the Eastern Cape province, South Africa. *South African Journal of Science*. 2002; 98: 589-597.
- [28] Okole BN and Odhav B. Commercialisation of plants in Africa. *South African Journal of Botany*. 2004; 70: 109–115.
- [29] Ndawonde BG. Medicinal plant sales: A case study in northern Zululand. MSc Dissertation. University of Zululand, KwaDlangezwa; 2006.
- [30] Monakisi CM. Knowledge and use of traditional medicinal plants by the Setswana-speaking community of Kimberley, Northern Cape of South Africa. MSc Dissertation. University of Stellenbosch, Stellenbosch; 2007.
- [31] Ndawonde BG, Zobolo AM, Dlamini ET and Siebert SJ. A survey of plants sold by traders at Zululand muthi markets, with a view to selecting popular plant species for propagation in communal gardens. *African Journal of Range and Forage Science*. 2007; 24: 103–107.
- [32] Williams VL, Witkowski ETF and Balkwill K. Volume and financial value of species traded in the medicinal plant markets of Gauteng, South Africa. *International Journal of Sustainable Development and World Ecology*. 2007; 14: 584-603.
- [33] Loundou PM. Medicinal plant trade and opportunities for sustainable management in the Cape Peninsula, South Africa. MSc Dissertation. University of Stellenbosch, Stellenbosch; 2008.
- [34] Ah Goo DFS. The contribution of the trade in medicinal plants to urban livelihoods: A case study of the informal markets in Nelson Mandela Bay Municipality, Eastern Cape. MSc Dissertation, Nelson Mandela Metropolitan University, Port Elizabeth; 2012.
- [35] Williams VL, Victor JE and Crouch NR. Red Listed medicinal plants of South Africa: Status, trends, and assessment challenges. *South African Journal of Botany*. 2013; 86: 23–35.
- [36] Beinart W and Beinart R. From elephant's foot...to cortisone': Boots Pure Drug Company and *Dioscorea sylvatica* in South Africa, c. 1950–1963. *South African Historical Journal*. 2019; 71: 644-675.
- [37] Lekganyane DM. The mystery behind muthi: A survey of bulbous and perennial herbs traded at the Faraday Medicinal Market in Johannesburg using DNA barcoding as an identification tool. MSc Dissertation. University Of Johannesburg, Johannesburg; 2019.
- [38] Rasethe MT, Semenya SS and Maroyi A. Medicinal plants traded in informal herbal medicine markets of the Limpopo province, South Africa. *Evidence-Based Complementary*

- and Alternative Medicine. 2019, volume 2019, article ID 2609532.
- [39] Wiersum KF, Husselman M, Dold AP and Cocks ML. Cultivation of medicinal plants as a tool for biodiversity conservation and poverty alleviation in the Amatola region, South Africa. In: Bogers RJ, Craker LE and Lange D (Eds.), *Medicinal and aromatic plants: Agricultural, commercial, ecological, legal, pharmacological and social aspects*. Springer, Dordrecht; 2006, pp. 43–57.
- [40] Nzue APMM. Use and conservation status of medicinal plants in the Cape Peninsula, Western Cape Province of South Africa. MSc Dissertation. University of Stellenbosch, Stellenbosch; 2009.
- [41] Philander LA. An ethnobotany of Western Cape Rasta bush medicine. *Journal of Ethnopharmacology*. 2011; 138: 578–594.
- [42] Xaba VM. Pharmacological Screening of Traditional Medicinal Plants used Against Skin Ailments in the Free State, South Africa. MSc Dissertation. University of the Free State, Phuthaditjhaba; 2016.
- [43] Hutchings A, Scott AH, Lewis G and Cunningham A. Zulu medicinal plants: An inventory. University of Natal Press, Pietermaritzburg; 1996.
- [44] Jolles F and Jolles S. Zulu ritual immunisation in perspective. *Africa Journal of the International African Institute*. 2000; 70: 229-248.
- [45] Rasethe MT. The utilization and management of selected listed-threatened or protected species in the Limpopo province, South Africa. MSc Dissertation. University of Limpopo, Sovenga; 2017
- [46] Long C. Swaziland's Flora: siSwati names and uses. Swaziland National Trust Commission; 2005. Available at: <http://www.sntc.org.sz/index.asp>, accessed on 15 February 2021.
- [47] Madikane EV. The isolation of anti-mycobacterial compounds from South African medicinal plants. PhD Thesis, University of Cape Town, Cape Town; 2005.
- [48] Van Wyk BV and Gericke N. *People's plants: A guide to useful plants of southern Africa*. Briza Publications, Pretoria; 2018.
- [49] Mbanjwa SG. A quantitative ethnobotanical survey of the Ixopo area of KwaZulu-Natal, South Africa. MSc Dissertation. University Of Johannesburg, Johannesburg; 2020.
- [50] Cocks ML and Wiersum KF. The significance of plant diversity to rural households in Eastern Cape province of South Africa. *Forests, Trees and Livelihoods*. 2003; 13: 39-58.
- [51] Cocks ML and Dold AP. Cultural significance of biodiversity: The role of medicinal plants in urban African cultural practices in the Eastern Cape, South Africa. *Journal of Ethnobiology*. 2006; 26: 60–81.
- [52] Street RA. Heavy metals in South African medicinal plants. PhD Thesis. University of KwaZulu-Natal, Pietermaritzburg; 2008.
- [53] Gelfand M, Mavi S, Drummond RB and Ndemera B. The traditional medical practitioner in Zimbabwe: His principles of practice and pharmacopoeia. Mambo Press, Gweru, Zimbabwe; 1985.
- [54] Semanya SS, Potgieter MJ and Erasmus LJC. Indigenous plant species used by Bapedi healers to treat sexually transmitted infections: Their distribution, harvesting, conservation and threats. *South African Journal of Botany*. 2013; 87: 66–75.
- [55] Erasmus LJC, Potgieter MJ, Semanya SS and Lennox SJ. Phytomedicine versus gonorrhoea: The Bapedi experience. *African Journal of Traditional Complementary and Alternative Medicine*. 2012; 9: 591- 598.
- [56] Semanya SS, Potgieter MJ and Erasmus LJC. Bapedi phytomedicine and their use in the treatment of sexually transmitted infections in Limpopo Province, South Africa. *African Journal of Pharmacy and Pharmacology*. 2013; 7: 250-262.
- [57] Komoreng L, Thekiso O, Lehasa S, Tiwani T, Mzizi N, Mokoena N, Khambule N, Ndebele S and Mdletshe N. An ethnobotanical survey of traditional medicinal plants used against lymphatic filariasis in South Africa. *South African Journal of Botany*. 2017; 111: 12–16.
- [58] Seaman T. The antimicrobial and antimycobacterial activity of plants used for the treatment of respiratory ailments in southern Africa and the isolation of anacardic acid from *Ozoroa paniculosa*. MSc Dissertation. Witwatersrand University, Johannesburg, 2005.
- [59] McGaw LJ, Lall N, Meyer JJM and Eloff JN. The potential of South African plants against *Mycobacterium* infections. *Journal of Ethnopharmacology*. 2008; 119: 482–500.
- [60] Chingwaru W, Vidmar J and Kapewangolo PT. The potential of sub-Saharan African plants in the management of human immunodeficiency virus infections: A review. *Phytotherapy Research*. 2015; 29: 1452–1487.
- [61] Semanya SS and Maroyi A. Data on medicinal plants used to treat respiratory infections and related symptoms in South Africa. *Data in Brief*. 2018; 21: 419-423.
- [62] Semanya SS and Maroyi A. Plants used by Bapedi traditional healers to treat asthma and related symptoms in Limpopo province, South Africa. *Evidence-Based Complementary and Alternative Medicine*. 2018; volume 2018, article ID 2183705.
- [63] Semanya SS and Maroyi A. Medicinal applications of plants by Bapedi traditional healers for sore throat and related symptoms in the Limpopo Province, South Africa. *Medicinal Plants: International Journal of Phytomedicines and Related Industries*. 2018; 10: 261-280.
- [64] Semanya SS and Maroyi A. Ethnobotanical survey of plants used by Bapedi traditional healers to treat tuberculosis and its opportunistic infections in the Limpopo province, South Africa. *South African Journal of Botany*. 2019; 122: 401–421.
- [65] Semanya SS and Maroyi A. Source of plants, used by Bapedi traditional healers for respiratory infections and related symptoms in the Limpopo Province, South Africa. *Journal of Biological Sciences*. 2019; 19: 101-121.
- [66] Semanya SS and Maroyi A. Source, harvesting, conservation status, threats and management of indigenous plant used for respiratory infections and related symptoms in the Limpopo Province, South Africa. *Biodiversitas*. 2019; 20: 790-811.
- [67] Semanya SS and Maroyi A. Ethnobotanical survey of plants used to treat respiratory infections and related symptoms in the Limpopo Province, South Africa. *Journal of Herbal Medicine*. 2020; 24: 100390.
- [68] Mavi S. Medicinal plants and their uses in Zimbabwe. In: Norman H, Snyman I and Cohen M (Eds.), *Indigenous knowledge and its uses in southern Africa*. Human Sciences Research Council, Pretoria; 1996, pp. 67-73.

- [69] Cogne A-L, Marston A, Mavi S and Hostettmann K. Study of two plants used in traditional medicine in Zimbabwe for skin problems and rheumatism: *Dioscorea sylvatica* and *Urginea altissima*. *Journal of Ethnopharmacology*. 2001; 75: 51–53.
- [70] Cogne A-L. Phytochemical investigation of plants used in African traditional medicine: *Dioscorea sylvatica* (Dioscoreaceae), *Urginea altissima* (Liliaceae), *Jamesbrittenia fodina* and *Jamesbrittenia elegantissima* (Scrophulariaceae). PhD Thesis. University of Lausanne, Lausanne; 2002.
- [71] Amusan OOG. Ethical and environmental issues in bioprospecting for drugs through traditional medicine: The case of Swaziland. *The African Journal of Plant Science and Biotechnology*. 2008; 2: 1-9.
- [72] Twilley D and Lall N. African plants with dermatological and ocular relevance. In: Kuete V (Ed.), *Toxicological survey of African Medicinal Plants*. Elsevier, London; 2014, pp. 493-512.
- [73] Xaba VM and Buwa LV. Pharmacological screening of traditional medicinal plants used to treat skin ailments in the Free State Province of South Africa. *South African Journal of Botany*. 2016; 103: 355.
- [74] Mwinga JL, Makhaga NS, Aremu AO and Otang-Mbeng W. Botanicals used for cosmetic purposes by Xhosa women in the Eastern Cape, South Africa. *South African Journal of Botany*. 2019; 126: 4–10.
- [75] Mander W, Mander J, Crouch N, McKean S and Nichols G. Catchment action: Growing and knowing muti plants. Institute of Natural Resources, South Africa ShareNet Booklet, Durban; 1995.
- [76] Rabe T and Van Staden J. Antibacterial activity of South African plants used for medicinal purposes. *Journal of Ethnopharmacology*. 1997; 56: 81–87.
- [77] Kelmanson JE, Jäger AK and Van Staden J. Zulu medicinal plants with antibacterial activity. *Journal of Ethnopharmacology*. 2000; 69: 241–246.
- [78] Wintola OA and Afolayan AJ. Ethnobotanical survey of plants used for the treatment of constipation within Nkonkobe Municipality of South Africa. *African Journal of Biotechnology*. 2010; 9: 7767-7770.
- [79] Watt JM and Breyer-Brandwijk MG. *The medicinal and poisonous plants of southern and eastern Africa*. 2nd ed. Livingstone, London; 1962.
- [80] Steyn DG. An investigation into cases of suspected poisoning in Africans in Northern Rhodesia. *South African Medical Journal*. 1965; 39: 344-350.
- [81] McGaw LJ and Eloff JN. Ethnoveterinary use of southern African plants and scientific evaluation of their medicinal properties. *Journal of Ethnopharmacology*. 2008; 119: 559–574.
- [82] Hossen MJ, Uddin MB, Ahmed SSU, Yu ZL and Cho JY. Traditional medicine/plants for the treatment of reproductive disorders in Asia nations. *Pakistan Veterinary Journal*. 2016; 36: 127-133.
- [83] Moore BD, Isiddro E and Seemann JR. Distribution of 2-carboxyarabinitol among plants. *Phytochemistry*. 1993; 34: 703-707.
- [84] Sibandze GF, Stapleton P and Gibbons S. Efflux inhibitors from Swazi medicinal plants. *Planta Medica*. 2016; 82: 381.
- [85] Sibandze GF, Stapleton P and Gibbons S. Constituents of two *Dioscorea* species that potentiate antibiotic activity against MRSA. *Journal of Natural Products*. 2020; 83: 1696–1700.
- [86] Blunden G and Hardman R. Thin-layer chromatography of *Dioscorea* sapogenins. *Journal of Chromatography*. 1964; 15: 273-276.
- [87] Martin FW. The species of *Dioscorea* containing sapogenin. *Economic Botany*. 1969; 23: 373-379.
- [88] Lehasa SG, Pieters R, Thekisoe MM and Komoreng LV. Biological activity of traditional medicinal plants used against lymphatic filariasis in the eastern Free State. *South African Journal of Botany*. 2017; 109: 344.